

## 2.5 Endocrine disruptors and thyroid-related disorders and diseases

### 2.5.1 Overview of thyroid-related disorders and diseases in humans and wildlife and evidence for endocrine disruption

Thyroid diseases and disorders in humans (e.g. congenital hypothyroidism and adult autoimmune thyroid disease) have increased in incidence over the past several decades, such that the burden of thyroid disease is approximately two billion people worldwide.

- **Thyroid diseases and disorders represent a particularly high and increasing disease burden in children and adolescents** in several countries in which they have been studied (McGrogan et al., 2008).
- **Between 6-10% of adults have a thyroid disease or disorder.** Hypothyroidism is the most common thyroid disorder and is six times more common in women than men (Vanderpump, Turnbridge & French, 1995)
- **Population-wide testing of thyroid function in the absence of suggestive clinical features reveals a great proportion of “mild” thyroid abnormalities that have most likely gone unrecognized.** These studies suggest that there may be many more adults with undiagnosed thyroid conditions than is currently appreciated (e.g. Canaris et al., 2000).
- Slight decreases in thyroid function - sometimes referred to as “subclinical” or mild hypothyroidism - may have adverse health consequences (elevated cholesterol levels, heart disease and diabetes), especially over the long term and during pregnancy.

Both genetic and environmental factors play a role in thyroid health. However, observations in laboratory animals and wildlife suggest that exposure to endocrine disruptors, particularly during fetal life, could also play a role. Alongside the human health trends, studies describing thyroid dysfunctions in wildlife also exist. Sometimes, these wildlife observations are associated with exposures to contaminants. Examples include:

- **Relationships between body burden of persistent organic pollutants (PCBs, PBDEs and organochlorine pesticides) and thyroid-related effects in marine mammals;** in seals (Brouwer., 1989; Hall, Kalantzi & Thomas, 2003; Hall & Thomas, 2007; Routti et al., 2008), sea lions (Debiec et al., 2005), beluga whales inhabiting the St. Lawrence estuary (DeGuise et al., 1995), the harbour porpoise (Schnitzler et al., 2008), and polar bear (Braathen et al., 2004; Skaare et al., 2001).

- **Significant thyroid disruption in monitoring studies of birds in the Great Lakes, Barents Sea, Tokyo Bay, linked with EDC (PBDE and PCB) burdens** (Scanes & McNabb, 2003; Verreault et al., 2004; Saita et al., 2004).
- **Thyroid disruption in salmonid fish living in heavily polluted regions of the Great Lakes in the United States during the 1970s and 1980s and, more recently, in mummichogs in New Jersey and San Francisco Bay** (reviewed in Jobling & Tyler, 2003; Zhou et al., 2000; Brar et al., 2010). Effects in mummichogs were positively correlated with PCB concentrations measured in the livers of the fish.

#### Hormonal mechanisms underlying thyroid disorders and diseases

The thyroid gland is located at the base of the throat and straddles the trachea. When it becomes physically enlarged in some diseases, it is visible to the eye or can be palpated (goitre). The major product of the thyroid gland is “thyroxine” (tetraiodothyronine,  $T_4$ ). However,  $T_4$  is not considered to be the most active form of the hormone; rather, it is converted to tri-iodothyronine ( $T_3$ ), which then acts on the thyroid hormone receptor (TR) in cells.

Thyroid function itself is controlled by “Thyroid-Stimulating Hormone” (TSH, or “thyrotropin”). TSH is a large protein hormone secreted from the pituitary gland that binds to specific membrane receptors on thyroid cells and activates a biochemical pathway that stimulates thyroid hormone production and secretion (Taurog, 2004). The amount of TSH stimulation required to maintain blood levels of thyroid hormone within a “normal” range is controlled by a negative feedback relationship between serum  $T_4$  and serum TSH (Larsen, Silva & Kaplan, 1981). The negative feedback action of  $T_4$  occurs both at the level of the hypothalamus (Vella & Hollenberg 2009; Hollenberg 2008; Greer et al., 1993; Koller et al., 1987; Aizawa & Greer 1981) and pituitary (Wan, Farhoud & Privalsky, 2005; Hodin et al., 1989; Chin & Carr, 1987; Carr, Need & Chin, 1987). Thus, under normal conditions, there is a negative correlation between serum levels of  $T_4$  (specifically “free”  $T_4$ ) and serum TSH.

For this reason, blood levels of  $T_4$  and TSH form the principle clinical measures of thyroid function and disease. So-called “reference” ranges are developed for human populations because there are slight differences in the set-point around which thyroid hormone is regulated in different races, ethnicities and in pregnancy. These reference ranges are generated from a large sample of the population that is without other measures of thyroid disease (symptoms or the presence of anti-thyroid antibodies) (Haddow et al., 2004; Surks, 1991). Reference ranges have been developed for different populations (e.g. Zarkovic et al., 2011), for the different periods of pregnancy (Haddow et al., 2004), even for twin versus singleton pregnancy (Haddow, Palomaki & McClain, 2006), and for preterm versus term birth (Clark et al., 2001; Adams et al., 1995).

Thyroid hormones are important for normal development of the human brain (Bernal, 2007; 2011; Oerbeck et al., 2007), lungs (van Tuyl et al., 2004; Bizzarro & Gross, 2004), heart

(Stoykov et al., 2006; Grover, Mellstom & Malm, 2005; Danzi, Dubon & Klein, 2005), and other organs. Moreover thyroid hormones induce metamorphosis in some fish (Yamano et al., 1994) and in frogs (Buchholz, Paul & Shi, 2005), and they are essential for development in birds (McNabb, 2006) and mammals (Zoeller & Rovet, 2004). There is remarkable evolutionary conservation among vertebrates and some invertebrates in the chemistry of thyroid hormones, as well as in their role in development and adult physiology (Heyland, Reitzel & Hodin, 2004; Heyland & Moroz, 2005). Likewise, the molecular signalling pathways (involving thyroid hormone receptors) through which these hormones exert their actions are highly conserved across the vertebrate taxa (Buchholz, Paul & Shi, 2005; Bertrand et al., 2004; Whitfield et al., 1999).

### Endocrine disruptors as risk factors in thyroid disease and dysfunction

Given the importance of thyroid hormone in human and wildlife physiology, and the life-long effects of thyroid dysfunction during development, it is reasonable to carefully consider the possibility that environmental chemicals may interfere with the ability of thyroid hormone to perform its functions. There is a very large list of environmental chemicals – mostly human-made – that can cause a reduction in circulating levels of thyroid hormone in experimental animals (Howdeshell, 2002; Brucker-Davis, 1998). Not all of these

produce goitre, although they all reduce serum concentrations of thyroid hormone. Moreover, more environmental chemicals are being identified that can interfere directly with the receptor for thyroid hormone (Zoeller, 2010) or with other processes controlling thyroid hormone action (Gilbert et al., 2011; see **Figure 2.9**).

### Thyroid hormone dependent mechanisms of nervous system development in animals and humans

Severe thyroid hormone deficiency produces severe brain damage (Chen & Hetzel, 2010) and moderate or even transient insufficiency can cause specific developmental defects in rodents (Auso et al., 2004; Crofton, 2004; Crofton et al., 2000; Goldey et al., 1995; Goodman and Gilbert, 2007), and in humans (Haddow et al., 1999; Kooistra et al., 2006; Oerbeck et al., 2003; 2007; Pop et al., 1995; 2003; Pop & Vulmsa, 2005). Small differences (~25%) in point estimates of maternal T<sub>4</sub> or TSH during the early fetal period are associated with adverse outcomes in humans (e.g. reduced IQ scores), even though hormone levels are not outside the population reference range (Haddow, Palomaki & Williams, 2002; Morreale de Escobar, Obregon & Escobar del Rey 2000). However, in a hallmark study by Bongers-Schokking et al. (2000), the Mental Development Index of children with congenital hypothyroidism was affected by the age of onset of treatment with thyroid hormone, rather than the specific serum free T<sub>4</sub> concentration after treatment. Thus, the degree of

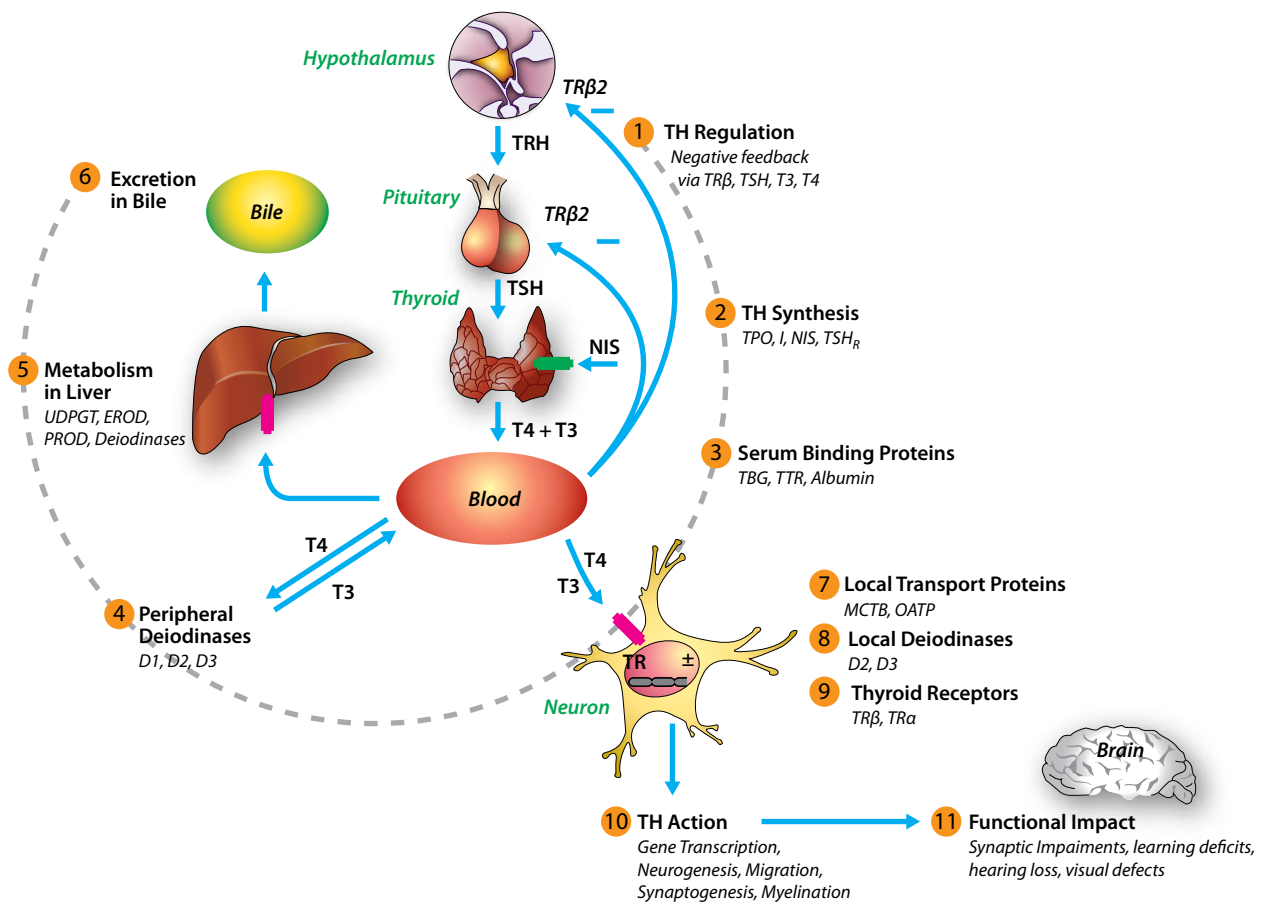


Figure 2.9. Possible sites of action of environmental contaminants on the HPT axis (Figure from Gilbert et al., 2011, redrawn; Used with publisher's permission).

thyroid hormone insufficiency is not the only variable affecting human development; the duration of the insufficiency and the developmental timing of the insufficiency are also important and may vary by species, presenting a challenge for risk assessment. This is discussed further in Chapter 1.2.4)

Experimental work in animals provides strong support for the hypothesis that moderate to mild thyroid hormone insufficiency can alter development in rodents. Integrating data over a series of studies, a decrease in serum total  $T_4$  by 50% during the critical period for cochlear development in the ear was associated with a permanent hearing loss in adult offspring (Crofton, 2004). Moreover, Auso et al. (2004) found that less than a 30% decrease in serum total  $T_4$  in female rodents, for only 3 days, was associated with structural abnormalities in the brains of their offspring. An average decrease in serum total  $T_4$  of only 28% in 2-week-old pups given low doses of propylthiouracil was associated with marked reduction in cell density of the corpus callosum region of the brain (Sharlin et al., 2008). Interestingly, Gilbert & Sui (2008) found that a 28% reduction in circulating levels of  $T_4$  in pregnant rats produced significant adverse effects on synaptic function of hippocampal neurons of their adult offspring despite no detected change in serum  $T_4$  levels in the pups after birth. The US EPA has discovered a cluster of neurons that reproducibly migrates to an incorrect position in the brain of animals that have low thyroid hormone (Goodman & Gilbert 2007). Elements of this cluster very sensitive to prenatal thyroid hormone insufficiency have been characterized (a heterotopia) (Gilbert et al., 2012). Finally, Sharlin et al., found a very strong inverse relationship between serum  $T_4$  in rat pups and the numbers of myelin-forming oligodendrocytes in major white matter tracks in the brain (Sharlin et al., 2008), and this was not compensated for by elevated serum TSH (Sharlin et al., 2010). Thus, experimental findings confirm what has been observed in humans: small, even transient, decreases in serum total  $T_4$  are associated with altered brain development.

In general, there is strong evidence to conclude that thyroid hormone plays the same general role in brain development of animals and humans (Zoeller & Rovet, 2004). This clearly indicates that rodents represent important test systems to provide information important for protecting public and wildlife health from chemical exposures. In animal studies, the investigator is able to measure the effect of environmental chemicals on blood levels of hormones, and can fully characterize the consequences of these changes on thyroid hormone action at the molecular, cellular and tissue level at various times during development. In addition, a variety of drugs and genetic lines of mice are available to experimentally confirm that environmental chemicals are specifically disrupting thyroid hormone action and not some other pathway of toxicity that could produce similar effects on apical endpoints. In contrast, in human studies, the investigator can only correlate measures of hormone levels in the blood with exposures and with various metrics of health and very few additional measures can be obtained to help interpret the relationship between these variables of interest. Therefore,

it is critically important to consider animal studies in the interpretation of human studies.

Notwithstanding this, the current set of validated test methods in the USA and EU for evaluating the ability of chemicals to interfere with thyroid hormone action does not include testing whether the chemical can interfere with thyroid hormone action (Zoeller, Tan & Tyl, 2007a; Zoeller, Tyl & Tan, 2007b).

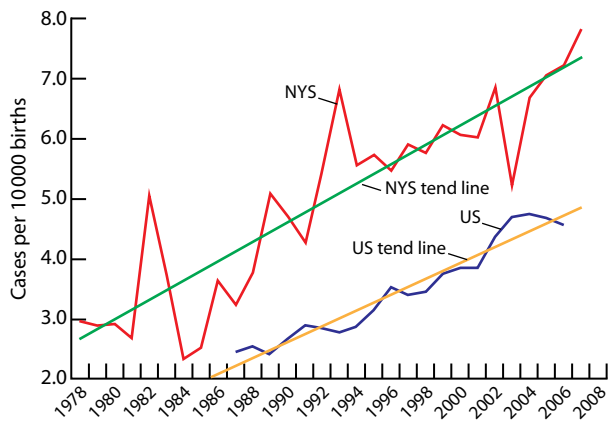
## 2.5.2 Evidence for endocrine disruption of the thyroid in humans and in mammalian models of humans

### 2.5.2.1 Human thyroid diseases and disorders

Thyroid disorders are amongst the most prevalent of medical conditions and include goitres or thyroid nodules (adults), congenital and adult hypothyroidism, autoimmune thyroiditis, hyperthyroidism or Graves' disease and thyroid cancer. In this section, we will deal mostly with congenital and adult hypothyroidism as well as Graves' disease, the remainder being covered in sections 2.11 (autoimmune diseases) and 2.7 (thyroid cancer). As already mentioned, thyroid hormone deficiencies during the development of the brain can also cause neurodevelopmental disturbances leading to mental difficulties, manifest as Attention Deficit Hyperactivity Disorder (ADHD), learning difficulties and possibly even autism. These are discussed further under section 2.6.

**Hypothyroidism:** This refers to an "underactive" thyroid gland such that it produces too little thyroid hormone. Symptoms associated with hypothyroidism are broad and can be somewhat non-specific including cold intolerance, weight gain, lethargy, and low mentation (Haddow, 2010). Moreover, the body consumes less oxygen and produces less body heat. Hypothyroidism can occur in both children and adults. In the adult population, studies in Northern Europe, Japan and the USA have found the prevalence of hypothyroidism to range between 0.6 and 12 per 1000 women and between 1.3 and 4.0 per 1000 men investigated, although the prevalence is higher in surveys of the elderly (Vanderpump, 2011).

**Congenital hypothyroidism:** Congenital hypothyroidism (CH) is one of the most common preventable causes of mental retardation caused by thyroid dysgenesis during fetal life. In the first trimester, the fetus is dependent on the trans-placental passage of thyroid hormones of maternal origin because the fetal thyroid gland does not produce thyroid hormone until the end of the first trimester, and then in sufficient quantities only at 20 weeks gestation (Smallridge et al., 2005). Thereafter, however, a hypothyroid fetus will synthesize around 70% less  $T_4$  than a normal fetus leading to CH (Olney, Grosse & Vogt, 2010). In 75-80% of all cases of CH, the underlying etiology is unknown, whilst the remaining 15-20% have genetic thyroid dysmorphogenesis. A daily iodine intake <25  $\mu\text{g}$ , particularly in preterm infants, accounts for many cases of CH in Europe, Asia and Africa, but multiple other factors may also be causal elements.



**Figure 2.10.** Incidence rate of CH in New York State (NYS), 1987–2007, and in the United States (excluding NYS), 1987–2006. (Figure from Hinton et al. (2010), redrawn; Used with publisher's permission).

Estimates of the birth prevalence of congenital hypothyroidism (CH) varies considerably throughout the world where universal screening programs are in place, as reviewed by Rendon-Macias et al. (2008). These estimates range from 1:1403 in Iran to 1:6450 in Latvia.

It was recently reported that the incidence of congenital hypothyroidism has nearly doubled over the past two decades in several countries in which it has been studied including the USA (Harris and Pass 2007; **Figure 2.10**), Western Australia (Kurinczuk et al., 2002), Italy (Corbetta et al., 2009), the northern UK (Pearce et al., 2010b), and Greece (Mengreli et al., 2010). Some authors speculate that this is due to changes in the cut-off values for the neonatal screening system in the definition of this disorder (Mitchell, Hso & Sahai, 2011; LaFranchi, 2011). This will be an important issue to address.

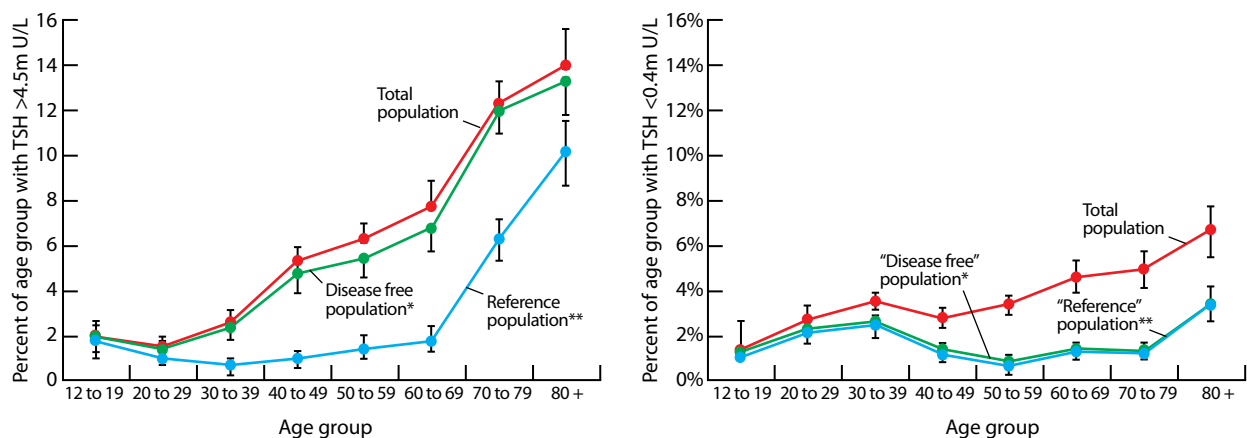
### Subclinical hypothyroidism (mild thyroid failure)

More widespread testing of thyroid function in the absence of suggestive clinical features suggests there are a great number of individuals, not diagnosed with thyroid problems, in which only TSH is abnormal (see **Figure 2.11**). A population study in Colorado, of over 25,000 individuals of mean age 56 years, showed TSH excess in 9.5% of the population and suppressed TSH in 2.2%; over half the group with suppressed TSH were taking thyroid medication. Similarly, in the Wickham survey in North East England, 8% of women and 3% of men had subclinical hypothyroidism and in the National Health and Nutrition Examination Survey (NHANES III), approximately 2% of adolescents aged 12–19 years had a serum TSH >4.5 mIU/L.

Prevalence data from one region do not necessarily apply to other populations, because of differences such as ethnic predisposition or variations in iodine intake. Several European studies have compared the effect of various levels of iodine intake on the prevalence of thyroid over- and under-function. Hypothyroidism is generally more common with abundant iodine intake, while goitre and subclinical hyperthyroidism are more common with low iodine intake.

### 2.5.2.2 Evidence for EDC exposures causing thyroid diseases and disorders.

It is possible that specific chemical exposures could lead to clinical thyroid disease and that this could be reflected in observed secular increases in the incidence or prevalence of thyroid disease. As reviewed above, thyroid disease is defined in large part by the presence of blood levels of  $T_4$  and TSH that are outside the population reference range. For example, clinical hypothyroidism is defined as low  $T_4$  and high TSH; both hormones need to be outside the reference range. However, the clinical symptoms associated with this hormone profile are highly variable in the population, and



**Figure 2.11.** Percentage of the USA population (in 2002) with abnormal serum TSH concentrations as a function of age. The disease-free population excludes those who reported thyroid disease, goiter or thyroid-related medications; the reference population excluded, in addition, those who had positive thyroid autoantibodies, or were taking medications that can influence thyroid function. Note the much higher prevalence of TSH abnormalities in the total population, than in the reference population (Figure from Hollowell et al. (2002), redrawn; Used with publisher's permission).

as a result, a significant proportion of the general population can have undiagnosed thyroid disease. In fact, in addition to the 14 million adults in the USA with diagnosed thyroid disease, a further 13 million are estimated to be undiagnosed (Blackwell, 2004). With such a large proportion of likely undiagnosed disease, it is clear that reported changes in incidence or prevalence would not be meaningful. Moreover, because thyroid hormone levels are variable within individuals (Andersen et al., 2003; Andersen et al., 2002), it will be difficult to identify relationships between clinical disease and chemical exposures; in contrast, it may be more likely that chemical exposures will be related to thyroid hormone levels within the reference range. Risk assessors should not disregard such relationships for several reasons:

- First, a large number of chemicals can affect circulating levels of thyroid hormone in animals (Howdeshell, 2002; Brucker-Davis, 1998). Although there are differences between rodents and humans in some characteristics of the thyroid system (see below), rodent systems still provide important fundamental information for the pharmaceutical development of therapeutics for humans. Therefore, it seems inefficient to employ rodent systems to develop drugs but to fail to use rodent systems to protect public health.
- Second, serum TSH levels within the reference range have been identified as a risk factor for blood pressure and serum cholesterol (reviewed in Miller et al., 2009) as well as for bone in postmenopausal women (Morris, 2007). This suggests that serum thyroid hormone levels – TSH and possibly total or free  $T_4$  – will be useful measures to link chemical exposures to various diseases.
- Finally, small differences in serum thyroid hormone levels during pregnancy or at birth are associated with deficits in cognitive function (LaFranchi, 2010). Therefore, if the fetus or neonate is as sensitive to chemical exposures as are adults, then even weak relationships between chemical exposure and hormone levels could produce permanent adverse effects.

A comprehensive review of this literature has recently appeared (Boas, Feldt-Rasmussen & Main, 2011; Boas, Main & Feldt-Rasmussen, 2009). There is now reasonably firm evidence that PCBs have thyroid-disrupting effects and that several other common contaminants also have such properties. These include brominated flame retardants, phthalates, bisphenol A and perfluorinated chemicals. In all cases, chemical exposure has been associated with serum thyroid hormone levels. Chemicals may affect circulating levels of thyroid hormone by interacting with the thyroid system in different ways (**Figure 2.9**) and there is currently little information about exactly how these may interact. A key issue is the extent to which changes in circulating levels of thyroid hormone reflect changes in thyroid hormone action in tissues (e.g. Zoeller, 2003). Human exposure to these chemicals (listed in Chapter 3, Table 3.1) is comprehensively reviewed in Chapter 3.2.2.

### 2.5.2.3 Polychlorinated biphenyls (PCBs)

PCBs are a family of biphenyls that have been randomly chlorinated, producing a mixture of chemicals that have as many as 209 different chlorination patterns. Their biological activity is altered by these patterns; in general, chlorination patterns that stabilize the ring structures into a planar conformation have dioxin-like activity (Kafafi et al., 1993; Kafafi et al., 1992) and those whose chlorination pattern stabilizes the ring structures into a non-coplanar conformation have a variety of activities (Lyng, 2007; Zoeller, 2001; Seegal & Shain, Snyder-Keller & Seegal, 1992; Shain, Bush & Seegal, 1991). Although PCB production was banned in the 1970s, PCBs remain common contaminants in the environment and in humans and wildlife both because of their chemical stability and because of the widespread use from heavy industrial applications to home products such as floor finishes and window caulking.

A number of studies have reported associations between PCB exposure and measures of thyroid function in humans that support the hypothesis that PCBs can reduce circulating levels of thyroid hormone (Abdelouahab et al., 2008; Hagmar et al., 2001a; 2001b; Persky et al., 2001; Schell et al., 2008; Turyk, Anderson & Persky, 2007). Some studies indicate that PCB body burdens suppress serum  $T_4$ , whilst others indicate serum  $T_3$ . In some cases, the findings are in men, in other cases in women. Overall, it is not a uniform picture. In studies of pregnant women, PCB body burden is positively associated with serum TSH (Chevrier et al., 2007; Takser et al., 2005). Studies of newborns also indicate that PCB body burden suppresses thyroid function (Chevrier et al., 2007; Herbstman et al., 2008). However, a number of studies report no associations between PCB body burden and measures of thyroid function (e.g. Dallaire et al., 2009; Dallaire et al., 2008; Longnecker et al., 2000).

There are a very large number of variables that must be considered to identify a relationship of interest between PCB exposures and measures of thyroid function. These include the fact that PCBs have a very long half-life in the human body and that there are many different PCB congeners that could influence thyroid function differently. There are also slightly different congener profiles in different populations. Measures of thyroid function are also variable across the population (serum total and free  $T_4$  and  $T_3$  and TSH) and this is exacerbated when time-of-day (with which thyroid hormone levels vary) is not standardized. Likewise, there are small gender and population differences. In one study of newborns, the birth mode (caesarean versus vaginal delivery) was important in identifying a relationship between serum PCBs and measures of thyroid function (Herbstman et al., 2008).

#### Evidence for PCB exposures causing thyroid diseases and disorders in rodent models

Considering these issues, it should be expected that not all studies will find exactly the same relationships. The issue is whether observed correlations between PCB body burden and various measures of thyroid function are consistent with an effect on population health that is mediated by effects on

thyroid hormone action. This is where experimental studies in animal models can be revealing. PCB exposures nearly uniformly cause a reduction in serum total and free  $T_4$  (Gauger, Sharlin & Zoeller, 2007a). However, serum TSH is not often reported to be elevated in response to this decrease (Hood and Klaassen). In addition, different PCB congeners appear to be differentially potent at causing serum  $T_4$  reductions (e.g. Giera et al., 2011), although it is not clear why this is observed. In a controlled study comparing the effects of reduced serum  $T_4$  produced by either propylthiouracil (PTU), which blocks thyroid hormone synthesis, or various PCBs, which presumably induce liver microsomes and decreases the serum half-life of  $T_4$ , Giera et al. (2011) found very different effects of PCB exposure compared to PTU exposure. Despite the fact that both exposures brought serum total  $T_4$  to the same concentration in blood, the two exposures had very different effects on the expression of known thyroid hormone response genes in the liver. Thus, the effect of PCB exposure on serum thyroid hormone levels cannot be interpreted the same way as the effect of PTU on serum thyroid hormone levels. This conclusion is supported by other studies (Bansal & Zoeller, 2008; Roegge et al., 2006; Bansal et al., 2005).

These findings also indicate that PCBs, or at least some congeners or metabolites, can interact directly with the thyroid hormone receptor. This hypothesis has been supported by a variety of studies. Several hydroxylated PCBs have been shown to displace  $T_3$  from the TR (You et al., 2006; Kitamura et al., 2005), or to increase (Freitas et al., 2011; Gauger et al., 2007) or decrease (Amano et al., 2010; Miyazaki et al., 2008) thyroid hormone receptor activation in expression systems. Likewise in vivo, PCBs produce effects that are consistent with the hypothesis that they can interfere with thyroid hormone action; in a recent study, PCB body burden in killer whales was highly correlated with the expression of the thyroid hormone receptor (Buckman et al., 2011), a known target of thyroid hormone itself.

Taken together, these findings reveal relatively inconsistent relationships between PCB exposure and measures of thyroid function in humans, but very strong evidence in animals and in molecular studies indicating that PCBs can interfere with thyroid hormone action. The complexity of the human data has been interpreted by some to indicate that there is no convincing evidence that PCBs interfere with thyroid function in humans (Kimbrough & Krouskas, 2003). Moreover, these authors suggest that even if the current data indicate that PCBs can interfere with thyroid function in humans, it is not clinically relevant. Importantly, this review did not include aspects of thyroid measurements that would provide insight into the difficulty in observing PCB effects of interest or the kind of statistical analysis that would be required.

All studies of endocrine disruptors in humans will likely have elements of the dataset observed with PCBs. Specifically, environmental chemicals may produce effects on endocrine systems that are either dissimilar to that of overt disease states, or that are inconsistent from one study to the next due to the difficulty in standardizing exposure measures and measures of hormone levels.

## 2.5.2.4 Other environmental chemicals

Boas, Feldt-Rasmussen and Main (2011) have also reviewed the literature linking a variety of chemical exposures to thyroid function in humans. These include PBDEs, pesticides, perfluorinated chemicals, phthalates, bisphenol A, UV-filters and perchlorate. With the possible exception of perchlorate, none of these chemicals have been as extensively for their relationship to thyroid function as that of PCBs. Human exposure to these chemicals is, however, extensive (Chapter 3.2.2). Suvorov and Takser (2008) suggest that the PCB story can further inform the number of publications (and time) required to generate enough data to make informed decisions about human and wildlife health.

## 2.5.2.5 The perchlorate controversy

Perchlorate is an oxidant used in a variety of industrial applications, from the production of solid rocket fuels, to explosives used in automobile airbags, fireworks and blasting caps (reviewed in Oxley et al., 2009). Perchlorate is also naturally occurring (Dasgupta et al., 2006), though the relative degree to which environmental contamination is caused by human-made or naturally occurring perchlorate is not clear. Perchlorate is chemically stable when wet and persists for long periods in geological systems and in groundwater. Largely because of disposal practices during the 1960s – 1990s, perchlorate became a common contaminant of groundwater in the United States (Urbansky, 2002).

The best known biological effect of perchlorate is the inhibition of iodide uptake by the sodium/iodide symporter (NIS) (Wolff, 1998), although it has recently been reported that perchlorate also interacts with Pendrin, another iodide transporter (Attanasio et al., 2011). NIS is responsible for transporting iodide into the thyroid gland, which is required for the production of thyroid hormone (Carrasco, 2000). In addition, this protein is expressed in the gut (Nicola et al., 2009; Vayre et al., 1999), lactating breast (Nicola et al., 2009; Dohan et al., 2003; 2007), placenta (Mitchell et al., 2001), and choroid plexus (Carrasco, 2000), all presumably as a delivery mechanism for iodide to the thyroid gland. In this regard, it is important that the expression of NIS in the human fetal thyroid gland is the limiting step in the production of thyroid hormone (Szinnai et al., 2007).

Given the essential role of thyroid hormone in development, it is important to determine whether perchlorate exposure is associated with measures of reduced thyroid function in the human population. Early studies sought to test this by comparing  $T_4$  or TSH levels in blood spots taken as part of the neonatal screening program with a proxy measure of perchlorate exposure – i.e. the city in which the infant was born (Las Vegas compared to Reno, Nevada, USA) (Li et al., 2000a; 2000b; Crump et al., 2000; Lamm and Doemland, 1999). The hypothesis was that because municipal drinking water was contaminated with perchlorate in Las Vegas but not in Reno, pregnant women and neonates would be exposed to perchlorate in Las Vegas but not in Reno. These studies uniformly found no association between the city of birth and neonatal thyroid

hormone. This was further supported by studies in Chile, in which perchlorate of natural origin is high, and again found no association between neonatal measures of thyroid function and the city of birth (Crump et al., 2000; Tellez Tellez, 2005). It was later shown in national biomonitoring data that almost everyone in the USA is exposed to perchlorate on a continual basis (Blount et al., 2006a; 2006b) and that much of this is derived from food (Huber et al., 2010; Sanchez et al., 2009). Therefore, studies using point estimates of exposure (i.e. city of birth) were confounded by large misclassifications of exposure and provide little useful information concerning the relationship of interest, i.e. perchlorate exposure and thyroid function.

A separate series of studies were performed to determine the efficacy of perchlorate exposure on iodide uptake inhibition in human volunteers (Greer et al., 2002; Lawrence, Lamm & Braverman, 2000; 2001), with the idea that this would help determine whether human exposures could influence thyroid function in the general population. These studies indicated that adults would have to consume 2L of drinking water daily that was contaminated with at least 200 ppb ( $\mu\text{g/L}$ ) perchlorate to reach a level in which iodide uptake would begin to be inhibited (Greer et al., 2002). Of course, the relationship between iodide uptake inhibition, thyroid hormone synthesis and serum concentrations of thyroid hormone is not known, but was believed to require significant iodide uptake inhibition for extended periods before thyroid function would be impaired. Based on these studies, a USA National Academy of Science (NAS) committee recommended a reference dose (RfD) of 0.0007 mg/kg per day (National Research Council, 2005), which the US EPA used to set a provisional drinking water standard of 15 ppb.

Several authors disagreed with EPA's drinking water standard of 15 ppb and perchlorate remediation goal of 24.5 ppb on the basis that it did not consider infants (Ginsberg et al., 2007). The reason for this was that infants must synthesise their supply of thyroid hormone each day (van den Hove et al., 1999); thus, if environmental factors reduce thyroid hormone synthesis and hormone levels decline, adverse effects on cognitive function would develop. Infants are very sensitive to thyroid hormone insufficiency (Zoeller & Rovet, 2004) and small differences in circulating levels of thyroid hormone in infants have been associated with differences in measures of cognitive function into adulthood (LaFranchi & Austin, 2007; Oerbeck et al., 2003; Heyerdahl & Oerbeck, 2003). Ginsberg et al. (2007) calculated that as many as 90% of nursing infants may exceed the RfD, although later empirical measurements indicate that this number is probably closer to 10% (Valentin-Blasini et al., 2011).

Blount et al. (2006b) showed a significant and sizable association between urinary perchlorate and serum thyroid hormones in a statistically representative sample of the USA population as part of the NHANES survey. This association was observed for women, but not for men. Importantly, the associations observed are plausibly consistent with a cause-effect relationship. That is, urinary perchlorate was positively associated with serum TSH and this association was stronger when urinary iodide was low. In addition, urinary perchlorate was negatively associated with serum  $T_4$  levels when urinary

iodide was low. Thirty percent of women in this study had low urinary iodide (below 100  $\mu\text{g/L}$ ). Using this same dataset, Steinmaus, Miller & Howd, (2007) showed that women who smoked had elevated levels of thiocyanate, which also inhibits iodide uptake by the NIS, and that in women with low urinary iodide, the association between perchlorate exposure and measures of thyroid function was much stronger.

Conclusions from the NHANES 2001-2002 data are not easily reconciled with the earlier studies of human volunteers or with other population studies (Pearce et al., 2010a; 2011). If these studies reflect a true relationship between very low levels of perchlorate exposure and thyroid function, it would mean that data derived from short-term, high-dose experiments in humans do not accurately predict effects of chronic low-dose exposures.

The conflicting findings among epidemiological studies of the relationship between perchlorate exposure and thyroid function should highlight features of the thyroid system that do not appear to be commonly taken into consideration. One of the most important of these is that circulating levels of thyroid hormone are somewhat variable in each individual (Andersen et al., 2002). In fact, Andersen et al. (2002) estimate that it would require 25 separate tests to estimate the "set point" for serum  $T_4$  in a single individual with a precision of 5%. Thus, the known variability in measurements of  $T_4$  and TSH should be employed to estimate the number of subjects needed to test whether there is a relationship between serum  $T_4$  and perchlorate. Likewise, consideration needs to be given to the known variability of estimates of perchlorate exposure. None of the current studies formally calculate the number of participants that would be required to identify a relationship between serum  $T_4$  (or TSH) and urinary perchlorate. The Blount study included over 1,111 women in their study – the largest to date.

The story of perchlorate contamination should be used to inform studies of other contaminants and their relationship with thyroid function. For those exposures that will act by changing circulating levels of thyroid hormone, perchlorate can serve as a direct example and it will be important to ensure that the study has enough subjects to provide adequate statistical power. This is important because there are known associations between circulating levels of thyroid hormone in pregnant women and, especially, neonates that provide very strong evidence linking hormone levels to adverse outcome. However, for exposures to chemicals that can interfere with thyroid hormone signalling without affecting serum hormone levels, there is clearly a lack of approach at this moment to test these associations in the human population.

### 2.5.3 Thyroid hormone and other organ systems

It is important to recognize that thyroid hormone concentrations are correlated with adverse effects in organ systems other than the nervous system in the adult, including the cardiovascular system and control of serum lipids (Asvold et al., 2007a; Biondi et al., 2005; Osman et al., 2002), pulmonary system (Krude et al., 2002; Lei et al., 2003; Mendelson & Boggaram, 1991) and

kidney. Total cholesterol, low density lipoproteins (LDL), non-high density lipoproteins (non-HDL), and triglycerides increase linearly with increasing TSH, and HDL decreases consistently with increasing TSH across normal reference ranges without evidence of any threshold effect (Asvold et al., 2007b). Similar trends in lipid profiles can be identified across clinical categories from hypothyroid to euthyroid to hyperthyroid individuals (Canaris et al., 2000). Within the reference ranges for TSH, there is a linear positive association between TSH and both systolic and diastolic blood pressure (Asvold et al., 2007b). Intimal medial thickness (IMT), a measure of atherosclerosis and predictive of coronary vascular disease and stroke, is inversely related to free  $T_4$  after controlling for lipids, clinical factors, and thyroid autoantibodies (Dullart et al., 2007). Some of these measures are ameliorated by treatment with thyroxine. Not surprisingly, deficits in thyroid homeostasis are associated with cardiovascular risk in multiple epidemiologic studies. A meta-analysis of 14 epidemiologic studies (Rodondi et al., 2006) found an overall increase in risk of coronary heart disease (CHD) of over 65% in those with subclinical hypothyroidism (elevation in TSH with normal  $T_4$ ). A higher risk was noted in those studies that adjusted for most cardiovascular risk factors. Treatment with L-thyroxine of patients with subclinical hypothyroidism resulted in improvements in cardiovascular risk factors including total cholesterol and endothelial function (Razvi et al., 2007). In addition, environmental exposure to at least one thyroid disrupting chemical (PCBs) has been shown to have an inverse association with  $T_3$  in men (Meeker, Altshu & Hauser, 2007) and was associated with both unfavorable lipid profiles and self reported cardiovascular disease in men and women (Goncharov et al., 2008). Therefore, epidemiologic as well as mechanistic and therapeutic evidence substantiates the concern that thyroid disrupting chemicals may adversely affect cardiovascular risk in humans by reducing serum  $T_4$ .

## 2.5.4 Evidence for endocrine disruption of the thyroid in vertebrate wildlife

Thyroid hormone is produced in all vertebrate classes and the chemistry of the hormone is identical in all of these species. In addition, thyroid hormones play a role in development in at least some members of all vertebrate classes. For example, in the flounder, metamorphosis is thyroid hormone dependent. This is also the case for amphibians. Much less is known about the capacity for thyroid dysfunction by EDCs in reptiles and in birds (with the exception of chick development, which provides an important developmental model). Thyroid hormone receptors (both  $TR\alpha$  and  $TR\beta$ ) are highly conserved among the vertebrates, suggesting that thyroid disruptors in any vertebrate may exert similar effects across all vertebrate species. However, metabolism of chemicals and subsequent exposures may differ considerably among the vertebrates and there may be other important differences that would suggest that caution be used when extrapolating information from one vertebrate class to another.

Thyroid hormone disruption reported in vertebrate wildlife species includes cetaceans and other sea mammals, as well

as a range of fish and birds. Some examples are given in the following sections. Effects on invertebrate wildlife have not been included: whilst thyroid hormone receptor orthologues have been reported across a range of invertebrate species, including the platyhelminths, *Schistosoma japonium* and *Schmidtea mediterranea*, the mollusc, *Lottia gigantea*, and the arthropod, *Daphnia pulex* (Wu, Niels & LoVerde, 2007), the capacity for thyrotoxic chemicals to exert effects on invertebrates is, as yet, unknown. Exposures of wildlife to thyroid hormone disrupting chemicals are comprehensively reviewed in Chapter 3.2.1.

### 2.5.4.1 Wild mammals

Many studies have reported relationships between individual body burdens of persistent organic pollutants and thyroid-related effects in seals (Brouwer, 1989; Hall, Kalantzi & Thomas, 2003; 2007; Routti et al., 2008), sea lions (Debieer et al., 2005), beluga whales inhabiting the St. Lawrence estuary (DeGuise et al., 1995), the harbour porpoise (Schnitzler et al., 2008), and the polar bear (Braathan et al., 2004; Skaare et al., 2001), suggesting contaminant-mediated disruption of thyroid homeostasis. In some studies, interfollicular fibrosis could be seen in the thyroid gland itself, associated with severe pathological dysfunction in other animals. PDBEs and PCBs particularly affect thyroid hormone transport and metabolism (Hallgren et al., 2001; Zhou et al., 2001; Zhou et al., 2002). Thyroid hormones are described as having a permissive role in the effects of other hormones and various enzymes, are important for metabolic regulation and are necessary for adequate growth. They control some aspects of fasting and may play a role in moulting cycles (Bentley et al., 1998). They are therefore key components of the endocrine system of wild mammals and any effects on their production, secretion, metabolism and target sites will have consequences for a range of physiological processes.

### 2.5.4.2 Non-mammalian vertebrates

Fish in contaminated locations are known to have impaired thyroid systems. The most famous historical examples of thyroid disruption were in the salmonids living in heavily polluted regions of the Great Lakes area in the United States during the 1970s and 1980s (e.g. Leatherland & Sontesgard, 1980a; 1980b; 1982a; 1982b; reviewed in Jobling & Tyler, 2003). Moreover, in the last decade, thyroid abnormalities were also reported in mummichogs from a polluted site in New Jersey, USA (Zhou et al., 2000) and in San Francisco Bay, California, USA (Brar et al., 2010). In the latter study, plasma concentrations of  $T_4$  were significantly reduced in two species of fish from highly contaminated areas, compared with fish from cleaner locations in the same estuary and both the  $T_3:T_4$  ratio and  $T_3$  concentrations were positively correlated with PCB concentrations measured in the livers of the exposed fish whilst  $T_4$  concentrations were inversely correlated. Taken together, the results support the conclusions from laboratory experiments and the general hypothesis already indicated in some marine and terrestrial mammals that environmental PCBs may alter  $T_4$  deiodination or turnover. Relationships between exposure to other chemicals



and thyroid hormone disruption in fish are less common, albeit increasing in the last decade, especially in relation to exposure to flame retardants (PBDEs).

In birds, biomarkers of exposure to thyroid-disrupting chemicals have also been evaluated by McNabb (2005), Panzica, Viglietti, Panzica & Ottinger (2005), and Grote et al. (2006). However, the exact extent to which EDCs exert effects on bird populations is still not established and field studies do not always support extrapolation from laboratory studies (e.g. Fernie et al., 2003; Fernie, Bortolotti & Smiths, 2003a; Fernie, Smiths & Bortolotti, 2003b), possibly because of between-species differences in susceptibility. Notwithstanding this, the relationships between the PCB concentrations and thyroid dysfunction in various bird species conducted over a long period strongly suggest that some PCBs can modulate this system in wild birds. This suggestion is now also supported by results from experimental studies on various model species. Long-term monitoring of herring gulls in the Great Lakes revealed significant thyroid dysfunction linked with PCB burden (Scanes & McNabb, 2003), and structural thyroid abnormalities detected in great cormorants from Tokyo Bay were also associated with PCDF and PCB contamination (Saita et al., 2004).

In addition, other studies on birds have found negative correlations between blood  $T_4$  and  $T_4:T_3$  ratio and levels of organochlorines, particularly hexachlorobenzene and oxychlorodane, in glaucous gulls from the Barents Sea (Verreault et al., 2004). Similarly, reduced  $T_4$  levels were reported in white stork nestlings exposed to pollution from a copper smelter (Kulczykowska et al., 2007). In contrast, an increase in  $T_3$  and  $T_4$  levels were detected within the thyroid glands of tree swallow nestlings from reclaimed wetlands partly filled with mine tailings from oil sands processing in Alberta, Canada (Gentes et al., 2007). It was postulated that the modulation of thyroid function in these birds may adversely affect metabolism, behaviour, feather development and moulting, ultimately compromising the survival of fledglings. High body burdens of PCBs in the European shag were associated with increased fluctuating wing asymmetry and also with disruption of the thyroid hormone, vitamin A (retinol) and vitamin E (tocopherol) homeostasis (Jenssen et al., 2010). Intergenerational effects of PCB exposure have also been demonstrated in kestrels, primarily via maternal transfer but also attributable to behavioural effects in the male parent. Where one or both parents had been exposed in ovo to PCBs, the progeny exhibited effects on development and growth, and sexually dimorphic effects on plasma  $T_3$  levels (Fernie et al., 2003b).

## 2.5.5 Evidence for a common EDC mechanism of thyroid disruption for human and wildlife

From the above, it is apparent that many of the symptoms associated with thyroid hormone disorders in humans, namely alterations in the levels of circulating thyroid hormones and

changes in the structure of the thyroid gland, have also been reported in wildlife. However, although probable, as yet there is no evidence that directly links the disruption of thyroid function via chemical exposure to adverse ecological effects in any wildlife species. In contrast, evidence of adverse effects is beginning to emerge from laboratory-based studies and will be discussed in the following section.

### 2.5.5.1 Evidence for EDC causation of thyroid disruption in laboratory studies with rodents and other vertebrates

Much of the laboratory-based research into the implications of EDC exposure for thyroid function in humans stems from studies using rodent models. For example, the rat has been extensively used to explore the health effects of exposure to PBDEs, with most studies consistently reporting a negative correlation with  $T_4$  concentrations (Zhou et al., 2002; Kodavanti & Derr-Yellin, 2002; Darnerud et al., 2007). Indeed, Kuriyama et al. (2007) demonstrated that BDE-99 has the capacity to reduce  $T_4$  levels in rats, even at low and environmentally relevant doses, with adipose tissue concentrations of BDE-99 in rats close to those reported in non-occupationally exposed humans and also at equivalent doses to those associated with other adverse outcomes in male and female rats, including permanent changes in neurobehaviour, locomotor activity and fertility (Kuriyama et al., 2005). Thus, it would appear that, in rodents, effects on thyroid function occur at EDC concentrations close to current human body burdens.

There is also laboratory-based evidence to support the assertion that EDCs are involved in the causation of thyroid disorders in wildlife species. For example, the suggestion that organochlorine pesticides, PCBs and flame retardants are causing thyroid disruption in arctic wildlife is supported by data from experimental studies on various model species such as domesticated arctic foxes, Greenland sled dogs and goats (e.g. Lyche et al., 2004; Oskam et al., 2004; Ropstad et al., 2006; Sonne et al., 2009). As a model of high trophic level carnivores, Kirkegaard et al. (2011) exposed female Greenland sled dogs and their pups to whale blubber contaminated with organohalogen compounds from 2-18 months of age and then examined thyroid hormone status. Although the sample numbers were low, the results supported observational data in other wildlife and humans, by showing that long term exposure to EDCs may result in detectable effects on thyroid hormone dynamics by lowering both free and total  $T_3$ .

In non-mammalian vertebrates, there are many laboratory studies reporting the effects of EDCs on thyroid hormone homeostasis, particularly in amphibians, due to the role of thyroid hormone in inducing metamorphosis. In this respect, BPA has been shown to block thyroid hormone-induced metamorphosis, indicating anti-thyroid activity (Iwamuro et al., 2003), which is consistent with its antagonism of  $T_3$  binding in *Xenopus* tadpoles (Goto et al., 2006). The herbicide acetochlor was also found to accelerate  $T_3$ -induced metamorphosis of *Xenopus* (Crump et al., 2002), a process that was preceded by disruption of  $T_3$ -

dependent expression of thyroid hormone receptor genes in the tadpole tail. Nonylphenol had an overall inhibitory effect on the rate of bullfrog tadpole metamorphosis (Christensen et al., 2005). Gutleb et al. (2007) developed a synchronized amphibian metamorphosis assay, which is based on the analysis of a range of endpoints, including the percentage of metamorphosed froglets by the end of the 60-day experimental period and the percentage of tadpoles at different stages of development, using *Xenopus laevis* as a model. Using this assay as a tool, a range of thyroid hormone disturbances were observed in response to a mixture of PCBs.

Although differences in sensitivity have been reported, depending on the model in question, in general, it would appear that the same chemicals, or groups of chemicals, elicit similar response patterns regardless of the species in question and the test system used. Laboratory-based studies using mammalian (mainly rodents) and non-mammalian species (most notably amphibians) have been invaluable in demonstrating the capacity for EDCs to affect thyroid development and in helping to identify critical periods of exposure during development. The data generated by these studies support the theory concerning the involvement of EDCs in the causation of thyroid disorders in wildlife and, in many cases, mirror the evidence concerning the etiology of these disorders in humans.

### 2.5.5.2 Interspecies extrapolation

Interspecies extrapolation of adverse effects of EDCs requires careful consideration. An example in which cross-species extrapolation is warranted is that of perchlorate. Perchlorate competitively inhibits iodine uptake into the thyroid gland, with subsequent decreases in TH synthesis and declines in circulating TH concentrations (Wolff, 1998). The kinetics for perchlorate inhibition of iodine uptake in humans and rats are extremely similar (US EPA, 2002), indicating the homologous nature of the initial toxic event. Although this is a clear example of a situation in which the toxic event (i.e. iodine uptake into the thyroid gland) exhibits similar kinetic profiles for rodents and humans, the impact of reduced serum thyroid hormone in rodents and humans may differ in some characteristics. For example, rodents or humans may possess robust compensatory mechanisms that would ameliorate the impacts of perchlorate exposure or low  $T_4$  (National Research Council, 2005). However, it is not at all clear that this is the case. Studies in humans indicate that even mild iodine insufficiency is associated with lower IQ in children (Berbel et al., 2009; Zimmermann, 2007; Aghini Lombardi et al., 1995), which does not support the notion that compensatory mechanisms are robust or available to the developing brain. Moreover in animals, Gilbert & Sui (2008) found that perchlorate exposure of pregnant rats can significantly affect synaptic transmission in the adult offspring, which also indicates that robust compensatory mechanisms to low thyroid hormone are not available. In addition, Sharlin et al. (2010) failed to identify compensatory responses to low levels of thyroid hormone in the developing rodent brain.

In contrast to the above, some studies do not support direct extrapolation between species (Crofton, 2004). To illustrate this kind of situation, both in vivo and in vitro studies suggest that PCBs activate the pregnane X receptor (PXR) in rodent liver, which leads to upregulation of hepatic catabolic enzymes and subsequent declines in circulating concentrations of  $T_4$  (Schuetz, Brimer & Schuetz, 1998). The steroid X receptor (SXR) is the human equivalent of rodent PXR (Blumberg et al., 1998) and there are species differences between these two proteins. Rodent PXR is activated by pregnenolone-16 $\alpha$ -carbonitrile (PCN), but not by rifampicin, whereas human SXR is activated by rifampicin but not by PCN (Kliewer, Goodwin & Willson, 2002). In addition, in vitro data suggest that high concentrations of CB-153 act as an antagonist at the human SXR rather than an agonist on the PXR in rodents (Tabb et al., 2004). Thus, PCBs may cause serum  $T_4$  to decline in animals but not in humans. While these data appear to support the conclusion that rodent data for PCBs are not relevant to humans, it does not appear to be that simple. First, if the hypothesis is correct that PCBs increase  $T_4$  clearance in a manner similar to that of phenobarbital, then serum TSH should increase as it does in response to phenobarbital (Hood & Klaassen, 2000). Because TSH does not increase in response to PCB exposure in rodents, the mechanism(s) by which PCBs cause a reduction in serum  $T_4$  may not be well understood. In addition, we know that some PCB congeners or metabolites can interact directly with the TR (see above), which is not related to a PXR/SXR pathway. Thus the mechanisms by which PCBs cause a reduction in serum  $T_4$  even in animals are not fully understood, nor have the most important pathways of toxicity in animals or humans been identified. Thus, the information required to exclude animal studies for consideration in risk assessment for PCBs is not available. Moreover, there are few other chemicals for which so much information is available. Therefore, it is unlikely to be the case that animal-to-human extrapolation should be excluded.

Finally, some authors propose that there are differences in circulatory transport proteins for thyroid hormones (e.g. transthyretin and thyroid-binding globulin) in rodents compared to humans and that this renders rodents much more sensitive to thyroid hormone reducing agents than are humans (Capen, 1997; Hill et al., 1998). However, it is not clear that these differences are meaningful for two reasons. First, pregnant and neonatal rodents have high levels of all transport proteins including thyroxine binding globulin (TBG) (Savu et al., 1991; Vranckx, Savu & Nunez, 1989; Savu et al., 1989; 1987). Rat TBG has been cloned (Tani et al., 1994) and its regulation studied (Vranckx et al., 1994). Thus, the contention that rodents do not have the same serum binding proteins as humans may not be correct. A further difference between rodents and humans is that the serum half-life of  $T_4$  in rodents is much shorter than that of humans (1 day in rodents versus 7-10 days in humans), although it is not at all clear that this issue renders rodent studies of thyroid function irrelevant to humans either; there are considerable data that suggest just the opposite.

## 2.5.6 Main messages

- Thyroid hormone is important in development and in adulthood in both wildlife and humans.
- Aside from thyroid cancer and congenital hypothyroidism, it is difficult to identify trends in the incidence of human thyroid disease.
- There are many chemicals that can interfere with thyroid function.
- Similarly, there are chemicals that can interfere directly with thyroid hormone action.
- Many chemicals interfere with thyroid function in a manner that will not be captured by evaluating only serum hormone levels.
- Despite the recognition that thyroid hormone is essential for brain development in humans, few if any chemicals are tested for their ability to interfere with thyroid hormone action.
- Relationships between exposure to chemicals and thyroid hormone disruption in wildlife species have increased in the last decade, especially in relation to exposure to flame retardants (PBDEs) and PCBs.
- The strength of evidence supporting a role for endocrine disrupting chemicals in disrupting thyroid function in wildlife adds credence to hypothesis that this could also occur in humans.
- Thyroid disruption is acknowledged to be poorly addressed by the chemical tests currently listed in the OECD Conceptual Framework.

## 2.5.7 Scientific progress since 2002

Since the *Global Assessment of the State-of-the-Science of Endocrine Disruptors* (IPCS, 2002), the following advances have been made:

- Increasing numbers of human studies establish a link between chemical exposures and thyroid function, including in pregnant women.
- However, few studies have focused on the relationship between chemical exposures in pregnant women, thyroid measures in those women (or in the cord blood of their offspring), and cognitive function in neonates.
- Genetic lines of mice have become widely available that should be coupled with toxicology studies to help clarify the mechanisms by which chemical exposures can interfere with thyroid hormone action.
- Relationships between exposure to chemicals and thyroid hormone disruption in wildlife species have increased in the last decade, especially in relation to exposure to the flame retardants (PBDEs) and PCBs, but other chemicals are insufficiently studied.

## 2.5.8 Strength of evidence

There is sufficient evidence that some thyroid diseases are increasing in the human population and that this may be related to environmental exposures. These diseases include congenital hypothyroidism and thyroid cancer. This evidence is considered to be sufficient because several authors report an increased incidence using screening data that reflect population-wide surveys. However, there are insufficient data linking these increases in thyroid disease to specific environmental factors.

There is limited evidence from wildlife studies and sufficient evidence from laboratory experiments that endocrine disrupting chemicals can interfere with thyroid hormone signalling, leading to diseases and disorders in wildlife species. The data generated by these studies support the theory concerning the involvement of EDCs in the causation of thyroid disorders in wildlife and mirror some of the evidence seen in humans. For many wildlife species, however, no studies have been done.

There is insufficient direct evidence in the human literature supporting the hypothesis that effects on thyroid hormone signalling mediate the association between chemical exposures and human disease/disorders. Perhaps the best example of this is focused on PCBs. There is sufficient evidence linking PCB body burden to reduced measures of cognitive function in children (Schantz, Widholm & Rice, 2003) and this evidence is deemed to be sufficient because a number of authors have reported similar findings and because it is consistent with studies in animals. In animal studies, PCBs clearly reduce circulating levels of thyroid hormone (Brouwer et al., 1998) and can affect brain development (Roegge et al., 2006). There are some studies indicating that PCB body burden is linked to reduced measures of cognitive function, but the evidence demonstrating a causal relationship is limited. Few studies have evaluated the relationship between PCB exposure, cognitive development, and thyroid hormone; therefore, there is overall insufficient evidence to demonstrate that PCBs interfere with thyroid hormone signalling and cause an adverse effect. Animal studies indicate that PCBs can exert effects on thyroid hormone signalling in development that are not consistent with effects on serum hormone levels (Bansal & Zoeller, 2008; Giera et al., 2011). Therefore, while considerable evidence exists in animal studies that chemicals can interfere with thyroid hormone signalling during development and produce adverse outcome, we have not developed the approach to fully test this hypothesis in human populations.

Thus, there are insufficient data linking chemical exposures to altered thyroid hormone signalling and the occurrence of disease or dysfunction in humans. Clearly, considering the importance of thyroid hormone during development, the large knowledge gaps, animal data, and the economic cost of population wide impacts on thyroid function during development (Dosiou et al., 2008), these are issues that need to be addressed quickly.

## 2.5.9 References

- Abdelouahab N, Mergler D, Takser L, Vanier C, St-Jean M, Baldwin M, Spear PA, Chan HM (2008). Gender differences in the effects of organochlorines, mercury, and lead on thyroid hormone levels in lakeside communities of Quebec (Canada). *Environmental Research*, 107(3):380-392.
- Adams LM, Emery JR, Clark SJ, Carlton EI, Nelson JC (1995). Reference ranges for newer thyroid function tests in premature infants. *The Journal of pediatrics*, 126(1):122-127.
- Aghini Lombardi FA, Pinchera A, Antonangeli L, Rago T, Chiovato L, Bargagna S, Bertucelli B, Ferretti G, Sbrana B, Marcheschi M, et al., (1995). Mild iodine deficiency during fetal/neonatal life and neuropsychological impairment in Tuscany. *Journal of Endocrinological Investigation*, 18(1):57-62.
- Aizawa T, Greer MA (1981). Delineation of the hypothalamic area controlling thyrotropin secretion in the rat. *Endocrinology*, 109:1731-1738.
- Amano I, Miyazaki W, Iwasaki T, Shimokawa N, Koibuchi N (2010). The effect of hydroxylated polychlorinated biphenyl (OH-PCB) on thyroid hormone receptor (TR)-mediated transcription through native-thyroid hormone response element (TRE). *Industrial Health*, 48(1):115-118.
- Andersen S, Pedersen KM, Bruun NH, Laurberg P (2002). Narrow individual variations in serum T(4) and T(3) in normal subjects: a clue to the understanding of subclinical thyroid disease. *Journal of Clinical Endocrinology and Metabolism*, 87(3):1068-1072.
- Andersen S, Bruun NH, Pedersen KM, Laurberg P (2003). Biologic variation is important for interpretation of thyroid function tests. *Thyroid*, 13(11):1069-1078.
- Asvold BO, Bjoro T, Nilsen TI, Vatten LJ (2007a). Association between blood pressure and serum thyroid-stimulating hormone concentration within the reference range: a population-based study. *Journal of Clinical Endocrinology and Metabolism*, 92(3):841-845.
- Asvold BO, Vatten LJ, Nilsen TI, Bjoro T (2007b). The association between TSH within the reference range and serum lipid concentrations in a population-based study. The HUNT Study. *European Journal of Endocrinology*, 156(2):181-186.
- Attanasio R, Scinicariello F, Blount BC, Valentin-Blasini L, Rogers KA, Nguyen DC, Murray HE (2011). Pendrin mediates uptake of perchlorate in a mammalian in vitro system. *Chemosphere*.
- Auso E, Lavado-Autric R, Cuevas E, Escobar Del Rey F, Morreale De Escobar G, Berbel P (2004). A moderate and transient deficiency of maternal thyroid function at the beginning of fetal neocortico-genesis alters neuronal migration. *Endocrinology*, 145(9):4037-4047.
- Bansal R, Zoeller RT (2008). Polychlorinated biphenyls (Aroclor 1254) do not uniformly produce agonist actions on thyroid hormone responses in the developing rat brain. *Endocrinology*, 149(8):4001-4008.
- Bansal R, You SH, Herzig CT, Zoeller RT (2005). Maternal thyroid hormone increases HES expression in the fetal rat brain: An effect mimicked by exposure to a mixture of polychlorinated biphenyls (PCBs). *Brain Research. Developmental Brain Research*, 156(1):13-22.
- Bentley PJ (1998). *Comparative vertebrate endocrinology*. Cambridge, UK ; New York, Cambridge University Press
- Berbel P, Mestre JL, Santamaria A, Palazon I, Franco A, Graells M, Gonzalez-Torga A, de Escobar GM (2009). Delayed neurobehavioral development in children born to pregnant women with mild hypothyroxinemia during the first month of gestation: the importance of early iodine supplementation. *Thyroid*, 19(5):511-519.
- Bernal J (2011). Thyroid hormone transport in developing brain. *Current Opinion in Endocrinology, Diabetes and Obesity*, 18(5):295-299.
- Bertrand S, Brunet FG, Escriva H, Parmentier G, Laudet V, Robinson-Rechavi M (2004). Evolutionary genomics of nuclear receptors: from twenty-five ancestral genes to derived endocrine systems. *Molecular Biology and Evolution*, 21(10):1923-1937.
- Biondi B, Palmieri EA, Klain M, Schlumberger M, Filetti S, Lombardi G (2005). Subclinical hyperthyroidism: clinical features and treatment options. *European Journal of Endocrinology*, 152(1):1-9.
- Bizzarro MJ, Gross I (2004). Effects of hormones on fetal lung development. *Obstetrics and Gynecology Clinics of North America*, 31(4):949-961, xii.
- Blackwell J (2004). Evaluation and treatment of hyperthyroidism and hypothyroidism. *Journal of the American Academy of Nurse Practitioners*, 16(10):422-425.
- Blount BC, Pirkle JL, Osterloh JD, Valentin-Blasini L, Caldwell KL (2006a). Urinary perchlorate and thyroid hormone levels in adolescent and adult men and women living in the United States. *Environmental Health Perspectives*, 114(12):1865-1871.
- Blount BC, Valentin-Blasini L, Osterloh JD, Mauldin JP, Pirkle JL (2006b). Perchlorate exposure of the US population, 2001-2002. *Journal of Exposure Science and Environmental Epidemiology*, 17(4):400-4007.
- Blumberg B, Sabbagh W, Jr., Juguilon H, Bolado J, Jr., van Meter CM, Ong ES, Evans RM (1998). SXR, a novel steroid and xenobiotic-sensing nuclear receptor. *Genes & Development*, 12(20):3195-3205.
- Boas M, Main KM, Feldt-Rasmussen U (2009). Environmental chemicals and thyroid function: an update. *Current Opinion in Endocrinology, Diabetes and Obesity*, 16(5):385-391.
- Boas M, Feldt-Rasmussen U, Main KM (2011). Thyroid effects of endocrine disrupting chemicals. *Molecular and Cellular Endocrinology*.
- Bongers-Schokking JJ, Koot HM, Wiersma D, Verkerk PH, de Muinck Keizer-Schrama SM (2000). Influence of timing and dose of thyroid hormone replacement on development in infants with congenital hypothyroidism. *The Journal of pediatrics*, 136(3):292-297.
- Braathen M, Derocher AE, Wiig O, Sormo EG, Lie E, Skaare JU, Jenssen BM (2004). Relationships between PCBs and thyroid hormones and retinol in female and male polar bears. *Environmental Health Perspectives*, 112(8):826-833.
- Brar NK, Waggoner C, Reyes JA, Fairey R, Kelley KM (2010). Evidence for thyroid endocrine disruption in wild fish in San Francisco Bay, California, USA. Relationships to contaminant exposures. *Aquatic Toxicology*, 96(3):203-215.
- Brouwer A (1989). Inhibition of thyroid hormone transport in plasma of rats by polychlorinated biphenyls. *Archives of toxicology. Supplement. = Archiv für Toxikologie. Supplement*, 13:440-445.
- Brouwer A, Morse DC, Lans MC, Schuur AG, Murk AJ, Klasson-Wehler E, Bergman A, Visser TJ (1998). Interactions of persistent environmental organohalogenes with the thyroid hormone system: mechanisms and possible consequences for animal and human health. *Toxicology and Industrial Health*, 14(1-2):59-84.
- Brucker-Davis F (1998). Effects of environmental synthetic chemicals on thyroid function. *Thyroid*, 8(9):827-856.
- Buchholz DR, Paul BD, Shi YB (2005). Gene-specific changes in promoter occupancy by thyroid hormone receptor during frog metamorphosis. Implications for developmental gene regulation. *Journal of Biological Chemistry*, 280(50):41222-41228.
- Buckman AH, Veldhoen N, Ellis G, Ford JK, Helbing CC, Ross PS (2011). PCB-associated changes in mRNA expression in Killer whales (Orcinus orca) from the NE Pacific Ocean. *Environmental Science and Technology*, 45(23):10194-10202.
- Canaris GJ, Manowitz NR, Mayor G, Ridgway EC (2000). The Colorado thyroid disease prevalence study. *Archives of Internal Medicine*, 160(4):526-534.
- Capen CC (1997). Mechanistic data and risk assessment of selected toxic end points of the thyroid gland. *Toxicologic Pathology*, 25(1):39-48.
- Carr FE, Need LR, Chin WW (1987). Isolation and characterization of the rat thyrotropin beta-subunit gene. Differential regulation of two

- transcriptional start sites by thyroid hormone. *Journal of Biological Chemistry*, 262(3):981-987.
- Carrasco N (2000). Throid iodide transport: the Na<sup>+</sup>/I<sup>-</sup> symporter (NIS). In: (Braverman LE, Utiger RD eds.) *The Thyroid: A Fundamental and Clinical Text*, Eighth edn., pp. 52-61. Philadelphia, Lippincott, Williams and Wilkins
- Chen ZP, Hetzel BS (2010). Cretinism revisited. *Best Practice and Research. Clinical Endocrinology and Metabolism*, 24(1):39-50.
- Chevrier J, Eskenazi B, Bradman A, Fenster L, Barr DB (2007). Associations between prenatal exposure to polychlorinated biphenyls and neonatal thyroid-stimulating hormone levels in a Mexican-American population, Salinas Valley, California. *Environmental Health Perspectives*, 115(10):1490-1496.
- Chin WW, Carr FE (1987). Thyroid hormone regulation of the rat thyrotropin beta-subunit gene. *Hormone and Metabolic Research. Supplement*, 17:82-86.
- Christensen JR, Richardson JS, Bishop CA, Pauli B, Elliott J (2005). Effects of nonylphenol on rates of tail resorption and metamorphosis in *Rana catesbeiana* tadpoles. *Journal of Toxicology and Environmental Health. Part A*, 68(7):557-572.
- Clark SJ, Deming DD, Emery JR, Adams LM, Carlton EI, Nelson JC (2001). Reference ranges for thyroid function tests in premature infants beyond the first week of life. *Journal of Perinatology*, 21(8):531-536.
- Corbetta C, Weber G, Cortinovis F, Calebiro D, Passoni A, Vigone MC, Beck-Peccoz P, Chiumello G, Persani L (2009). A 7-year experience with low blood TSH cutoff levels for neonatal screening reveals an unsuspected frequency of congenital hypothyroidism (CH). *Clinical Endocrinology*, 71(5):739-745.
- Crofton KM (2004). Developmental disruption of thyroid hormone: correlations with hearing dysfunction in rats. *Risk Analysis*, 24(6):1665-1671.
- Crofton KM, Ding D, Padich R, Taylor M, Henderson D (2000). Hearing loss following exposure during development to polychlorinated biphenyls: a cochlear site of action. *Hearing Research*, 144(1-2):196-204.
- Crump C, Michaud P, Tellez R, Reyes C, Gonzalez G, Montgomery EL, Crump KS, Lobo G, Becerra C, Gibbs JP (2000). Does perchlorate in drinking water affect thyroid function in newborns or school-age children? *Journal of Occupational and Environmental Medicine*, 42(6):603-612.
- Crump D, Werry K, Veldhoen N, Van Aggelen G, Helbing CC (2002). Exposure to the herbicide acetochlor alters thyroid hormone-dependent gene expression and metamorphosis in *Xenopus Laevis*. *Environmental Health Perspectives*, 110(12):1199-1205.
- Dallaire R, Dewailly E, Pereg D, Dery S, Ayotte P (2009). Thyroid function and plasma concentrations of polyhalogenated compounds in Inuit adults. *Environmental Health Perspectives*, 117(9):1380-1386.
- Dallaire R, Dewailly E, Ayotte P, Muckle G, Laliberte C, Bruneau S (2008). Effects of prenatal exposure to organochlorines on thyroid hormone status in newborns from two remote coastal regions in Quebec, Canada. *Environmental Research*, 108(3):387-392.
- Danzi S, Dubon P, Klein I (2005). Effect of serum triiodothyronine on regulation of cardiac gene expression: role of histone acetylation. *American Journal of Physiology. Heart and Circulatory Physiology*, 289(4):H1506-1511.
- Darnerud PO, Aune M, Larsson L, Hallgren S (2007). Plasma PBDE and thyroxine levels in rats exposed to Bromkal or BDE-47. *Chemosphere*, 67(9):S386-392.
- Dasgupta PK, Dyke JV, Kirk AB, Jackson WA (2006). Perchlorate in the United States. Analysis of relative source contributions to the food chain. *Environmental Science and Technology*, 40(21):6608-6614.
- Debiec C, Ylitalo GM, Weise M, Gulland F, Costa DP, Le Boeuf BJ, de Tillesse T, Larondelle Y (2005). PCBs and DDT in the serum of juvenile California sea lions: associations with vitamins A and E and thyroid hormones. *Environmental Pollution*, 134(2):323-332.
- DeGuise S, Martineau D, Beland P, Fournier M (1995). Possible mechanisms of action of environmental contaminants on St-Lawrence Beluga whales (*Delphinapterus-leucas*). *Environmental Health Perspectives*, 103:73-77.
- Dohan O, Portulano C, Basquin C, Reyna-Neyra A, Amzel LM, Carrasco N (2007). The Na<sup>+</sup>/I<sup>-</sup> symporter (NIS) mediates electroneutral active transport of the environmental pollutant perchlorate. *Proceedings of the National Academy of Sciences of the United States of America*, 104(51):20250-20255.
- Dohan O, De la Vieja A, Paroder V, Riedel C, Artani M, Reed M, Ginter CS, Carrasco N (2003). The sodium/iodide Symporter (NIS): characterization, regulation, and medical significance. *Endocrine Reviews*, 24(1):48-77.
- Dosiou C, Sanders GD, Araki SS, Crapo LM (2008). Screening pregnant women for autoimmune thyroid disease: a cost-effectiveness analysis. *European Journal of Endocrinology*, 158(6):841-851.
- Dullaart RP, de Vries R, Roozendaal C, Kobold AC, Sluiter WJ (2007). Carotid artery intima media thickness is inversely related to serum free thyroxine in euthyroid subjects. *Clinical Endocrinology*, 67(5):668-673.
- Fernie K, Bortolotti G, Smits J (2003a). Reproductive abnormalities, teratogenicity, and developmental problems in American kestrels (*Falco sparverius*) exposed to polychlorinated biphenyls. *Journal of Toxicology and Environmental Health. Part A*, 66(22):2089-2103.
- Fernie K, Smits J, Bortolotti G (2003b). Developmental toxicity of in ovo exposure to polychlorinated biphenyls: I. Immediate and subsequent effects on first-generation nestling American kestrels (*Falco sparverius*). *Environmental toxicology and chemistry / SETAC*, 22(3):554-560.
- Fernie K, Bortolotti G, Drouillard K, Smits J, Marchant T (2003). Developmental toxicity of in ovo exposure to polychlorinated biphenyls: II. Effects of maternal or paternal exposure on second-generation nestling american kestrels. *Environmental toxicology and chemistry / SETAC*, 22(11):2688-2694.
- Freitas J, Cano P, Craig-Veit C, Goodson ML, Furlow JD, Murk AJ (2011). Detection of thyroid hormone receptor disruptors by a novel stable in vitro reporter gene assay. *Toxicology in Vitro*, 25(1):257-266.
- Gaitan E (1989). Environmental goitrogenesis. CRC Press, Inc, Boca Raton, p. 250 pages.
- Gauger KJ, Sharlin DS, Zoeller RT (2007a). Polychlorinated biphenyls as disruptors of thyroid hormone action (Hansen L ed.). University of Illinois Press, Champagne-Urbana.
- Gauger KJ, Giera S, Sharlin DS, Bansal R, Iannacone E, Zoeller RT (2007b). Polychlorinated biphenyls 105 and 118 form thyroid hormone receptor agonists after cytochrome P4501A1 activation in rat pituitary GH3 cells. *Environmental Health Perspectives*, 115(11):1623-1630.
- Gentes ML, McNabb A, Waldner C, Smits JE (2007). Increased thyroid hormone levels in tree swallows (*Tachycineta bicolor*) on reclaimed wetlands of the athabasca oil sands. *Archives of Environmental Contamination and Toxicology*, 53(2):287-292.
- Giera S, Bansal R, Ortiz-Toro TM, Taub DG, Zoeller RT (2011). Individual polychlorinated biphenyl (PCB) congeners produce tissue- and gene-specific effects on thyroid hormone signaling during development. *Endocrinology*, 152(7):2909-2919.
- Gilbert ME, Sui L (2008). Developmental exposure to perchlorate alters synaptic transmission in hippocampus of the adult rat. *Environmental Health Perspectives*, 116(6):752-760.
- Gilbert ME, Rovet J, Chen ZP, Koibuchi N (2012). Developmental thyroid hormone disruption: Prevalence, environmental contaminants and neurodevelopmental consequences. *Neurotoxicology*, 33(4):842-852.
- Ginsberg GL, Hattis DB, Zoeller RT, Rice DC (2007). Evaluation of

- the U.S. EPA/OSWER preliminary remediation goal for perchlorate in groundwater: focus on exposure to nursing infants. *Environmental Health Perspectives*, 115(3):361-369.
- Goldey ES, Kehn LS, Rehnberg GL, Crofton KM (1995). Effects of developmental hypothyroidism on auditory and motor function in the rat. *Toxicology and Applied Pharmacology*, 135(1):67-76.
- Goncharov A, Haase RF, Santiago-Rivera A, Morse G, McCaffrey RJ, Rej R, Carpenter DO (2008). High serum PCBs are associated with elevation of serum lipids and cardiovascular disease in a Native American population. *Environmental Research*, 106(2):226-239.
- Goodman JH, Gilbert ME (2007). Modest Thyroid Hormone Insufficiency During Development Induces a Cellular Malformation in the Corpus Callosum: A Model of Cortical Dysplasia. *Endocrinology*, 148(6):2593-2597.
- Goto Y, Kitamura S, Kashiwagi K, Oofusa K, Tooi O, Yoshizato K, Sato J, Ohta S, Kashiwagi A (2006). Suppression of amphibian metamorphosis by bisphenol A and related chemical substances. *Journal of Health Science*, 52(2):160-168.
- Greer MA, Goodman G, Pleus RC, Greer SE (2002). Health effects assessment for environmental perchlorate contamination: the dose response for inhibition of thyroidal radioiodine uptake in humans. *Environmental Health Perspectives*, 110(9):927-937.
- Greer MA, Sato N, Wang X, Greer SE, McAdams S (1993). Evidence that the major physiological role of TRH in the hypothalamic paraventricular nuclei may be to regulate the set-point for thyroid hormone negative feedback on the pituitary thyrotroph. *Neuroendocrinology*, 57(4):569-575.
- Grote K, Niemann L, Gericke C, Selzsam B, Chahoud I (2006). Effects of fentin hydroxide on reproduction of the Japanese quail (*Coturnix coturnix japonica*). *Environmental Research*, 101(1):81-88.
- Grover GJ, Mellstrom K, Malm J (2005). Development of the thyroid hormone receptor beta-subtype agonist KB-141: A strategy for body weight reduction and lipid lowering with minimal cardiac side effects. *Cardiovascular drug reviews*, 23(2):133-148.
- Gutleb AC, Schriks M, Mossink L, van den Berg JHJ, Murk AJ (2007). A synchronized amphibian metamorphosis assay as an improved tool to detect thyroid hormone disturbance by endocrine disruptors and apolar sediment extracts. *Chemosphere*, 70(1):93-100.
- Haddow JE (2010). Hypothyroidism: detecting and treating early symptoms as the body's energy rheostat is slowly turned down. *Journal of Medical Screening*, 17(4):163.
- Haddow JE, Palomaki GE, Williams J (2002). Thyroid-stimulating-hormone concentrations and risk of hypothyroidism. *Lancet*, 360(9350):2081-2082; author reply 2082.
- Haddow JE, Palomaki GE, McClain MR (2006). Thyroid-stimulating hormone in singleton and twin pregnancy: importance of gestational age-specific reference ranges. *Obstetrics and Gynecology*, 107(1):205-206; author reply 206.
- Haddow JE, Knight GJ, Palomaki GE, McClain MR, Pulkkinen AJ (2004). The reference range and within-person variability of thyroid-stimulating hormone during the first and second trimesters of pregnancy. *Journal of Medical Screening*, 11(4):170-174.
- Haddow JE, Palomaki GE, Allan WC, Williams JR, Knight GJ, Gagnon J, O'Heir CE, Mitchell ML, Hermos RJ, Waisbren SE, Faix JD, Klein RZ (1999). Maternal thyroid deficiency during pregnancy and subsequent neuropsychological development of the child. *New England Journal of Medicine*, 341(8):549-555.
- Hagmar L, Bjork J, Sjodin A, Bergman A, Erfurth EM (2001a). Plasma levels of persistent organohalogen and hormone levels in adult male humans. *Archives of Environmental Health*, 56(2):138-143.
- Hagmar L, Rylander L, Dyremark E, Klasson-Wehler E, Erfurth EM (2001b). Plasma concentrations of persistent organochlorines in relation to thyrotropin and thyroid hormone levels in women. *International Archives of Occupational and Environmental Health*, 74(3):184-188.
- Hall AJ, Thomas GO (2007). Polychlorinated biphenyls, DDT, polybrominated diphenyl ethers, and organic pesticides in United Kingdom harbor seals (*Phoca vitulina*)--mixed exposures and thyroid homeostasis. *Environmental toxicology and chemistry / SETAC*, 26(5):851-861.
- Hall AJ, Kalantzi OI, Thomas GO (2003). Polybrominated diphenyl ethers (PBDEs) in grey seals during their first year of life--are they thyroid hormone endocrine disrupters? *Environmental Pollution*, 126(1):29-37.
- Hallgren S, Sinjari T, Hakansson H, Darnerud PO (2001). Effects of polybrominated diphenyl ethers (PBDEs) and polychlorinated biphenyls (PCBs) on thyroid hormone and vitamin A levels in rats and mice. *Archives of Toxicology*, 75(4):200-208.
- Harris KB, Pass KA (2007). Increase in congenital hypothyroidism in New York State and in the United States. *Molecular Genetics and Metabolism*, 91(3):268-277.
- Herbstman JB, Sjodin A, Apelberg BJ, Witter FR, Halden RU, Patterson DG, Panny SR, Needham LL, Goldman LR (2008). Birth delivery mode modifies the associations between prenatal polychlorinated biphenyl (PCB) and polybrominated diphenyl ether (PBDE) and neonatal thyroid hormone levels. *Environmental Health Perspectives*, 116(10):1376-1382.
- Heyerdahl S, Oerbeck B (2003). Congenital hypothyroidism: developmental outcome in relation to levothyroxine treatment variables. *Thyroid*, 13(11):1029-1038.
- Heyland A, Moroz LL (2005). Cross-kingdom hormonal signaling: an insight from thyroid hormone functions in marine larvae. *Journal of Experimental Biology*, 208(Pt 23):4355-4361.
- Heyland A, Reitzel AM, Hodin J (2004). Thyroid hormones determine developmental mode in sand dollars (Echinodermata: Echinoidea). *Evolution and Development*, 6(6):382-392.
- Hill RN, Crisp TM, Hurley PM, Rosenthal SL, Singh DV (1998). Risk assessment of thyroid follicular cell tumors. *Environmental Health Perspectives*, 106(8):447-457.
- Hinton CF, Harris KB, Borgfeld L, Drummond-Borg M, Eaton R, Lorey F, Therrell BL, Wallace J, Pass KA (2010). Trends in Incidence Rates of Congenital Hypothyroidism Related to Select Demographic Factors: Data From the United States, California, Massachusetts, New York, and Texas. *Pediatrics*, 125:S37-S47.
- Hodin RA, Lazar MA, Wintman BI, Darling DS, Chin WW (1989). Identification of a thyroid hormone receptor that is pituitary-specific. *Science*, 244:76-79.
- Hollenberg AN (2008). The role of the thyrotropin-releasing hormone (TRH) neuron as a metabolic sensor. *Thyroid*, 18(2):131-139.
- Hollowell JG, Staehling NW, Flanders WD, Hannon WH, Gunter EW, Spencer CA, Braverman LE (2002). Serum TSH, T4, and Thyroid Antibodies in the United States Population (1988 to 1994): National Health and Nutrition Examination Survey (NHANES III). *Journal of Clinical Endocrinology and Metabolism*, 87(2):489-499.
- Hood A, Klaassen CD (2000). Differential effects of microsomal enzyme inducers on in vitro thyroxine (T(4)) and triiodothyronine (T(3)) glucuronidation. *Toxicological sciences: an official journal of the Society of Toxicology*, 55(1):78-84.
- Howdeshell KL (2002). A model of the development of the brain as a construct of the thyroid system. *Environmental Health Perspectives*, 110 Suppl 3:337-348.
- Huber DR, Blount BC, Mage DT, Letkiewicz FJ, Kumar A, Allen RH (2010). Estimating perchlorate exposure from food and tap water based on US biomonitoring and occurrence data. *Journal of Exposure Science and Environmental Epidemiology*, 21(4):395-407.
- IPCS (2002) Global Assessment of the State-of-the-Science of

- Endocrine Disruptors. International Programme on Chemical Safety, World Health Organization, Geneva, Switzerland.
- Iwamuro S, Sakakibara M, Terao M, Ozawa A, Kurobe C, Shigeura T, Kato M, Kikuyama S (2003). Teratogenic and anti-metamorphic effects of bisphenol A on embryonic and larval *Xenopus laevis*. *General and Comparative Endocrinology*, 133(2):189-198.
- Jenssen BM, Aarnes JB, Murvoll KM, Herzke D, Nygard T (2010). Fluctuating wing asymmetry and hepatic concentrations of persistent organic pollutants are associated in European shag (*Phalacrocorax aristotelis*) chicks. *The Science of the total environment*, 408(3):578-585.
- Jobling S, Tyler CR (2003). Endocrine disruption in wild freshwater fish. *Pure and Applied Chemistry*, 75(11-12):2219-2234.
- Kafafi SA, Afeefy HY, Said HK, Hakimi JM (1992). A new structure-activity model for Ah receptor binding. Polychlorinated dibenzo-p-dioxins and dibenzofurans. *Chemical Research in Toxicology*, 5(6):856-862.
- Kafafi SA, Afeefy HY, Ali AH, Said HK, Abd-Elazem IS, Kafafi AG (1993). Affinities for the aryl hydrocarbon receptor, potencies as aryl hydrocarbon hydroxylase inducers and relative toxicities of polychlorinated biphenyls. A congener specific approach. *Carcinogenesis*, 14(10):2063-2071.
- Kimbrough RD, Krouskas CA (2003). Human exposure to polychlorinated biphenyls and health effects: a critical synopsis. *Toxicological Reviews*, 22(4):217-233.
- Kirkegaard M, Sonne C, Dietz R, Letcher RJ, Jensen AL, Hansen SS, Jenssen BM, Grandjean P (2011). Alterations in thyroid hormone status in Greenland sledge dogs exposed to whale blubber contaminated with organohalogen compounds. *Ecotoxicology and Environmental Safety*, 74(1):157-163.
- Kitamura S, Jinno N, Suzuki T, Sugihara K, Ohta S, Kuroki H, Fujimoto N (2005). Thyroid hormone-like and estrogenic activity of hydroxylated PCBs in cell culture. *Toxicology*, 208(3):377-387.
- Kliwer SA, Goodwin B, Willson TM (2002). The nuclear pregnane X receptor: a key regulator of xenobiotic metabolism. *Endocrine Reviews*, 23(5):687-702.
- Kodavanti PR, Derr-Yellin EC (2002). Differential effects of polybrominated diphenyl ethers and polychlorinated biphenyls on [3H] arachidonic acid release in rat cerebellar granule neurons. *Toxicological sciences : an official journal of the Society of Toxicology*, 68(2):451-457.
- Koller KJ, Wolff RS, Warden MK, Zoeller RT (1987). Thyroid hormones regulate levels of thyrotropin-releasing-hormone mRNA in the paraventricular nucleus. *Proceedings of the National Academy of Sciences of the United States of America*, 84(20):7329-7333.
- Kooistra L, Crawford S, van Baar AL, Brouwers EP, Pop VJ (2006). Neonatal effects of maternal hypothyroxinemia during early pregnancy. *Pediatrics*, 117(1):161-167.
- Krude H, Schutz B, Biebermann H, von Moers A, Schnabel D, Neitzel H, Tonnie H, Weise D, Lafferty A, Schwarz S, DeFelice M, von Deimling A, van Landeghem F, DiLauro R, Gruters A (2002). Choreaathetosis, hypothyroidism, and pulmonary alterations due to human NKX2-1 haploinsufficiency. *The Journal of clinical investigation*, 109(4):475-480.
- Kulczykowska E, Kasprzak M, Kalamaz H, Kuriata M, Nietrzeba M, Jerzak L, Kaminski P (2007). Melatonin and thyroxine response to pollution in white stork nestlings (*Ciconia ciconia*): Aspects of rhythmicity and age. *Comparative Biochemistry and Physiology, Part C: Toxicology & Pharmacology*, 146(3):392-397.
- Kurinczuk JJ, Bower C, Lewis B, Byrne G (2002). Congenital hypothyroidism in Western Australia 1981-1998. *Journal of Paediatrics and Child Health*, 38(2):187-191.
- Kuriyama SN, Talsness CE, Grote K, Chahoud I (2005). Developmental exposure to low dose PBDE 99: effects on male fertility and neurobehavior in rat offspring. *Environmental Health Perspectives*, 113(2):149-154.
- Kuriyama SN, Wanner A, Fidalgo-Neto AA, Talsness CE, Koerner W, Chahoud I (2007). Developmental exposure to low-dose PBDE-99: tissue distribution and thyroid hormone levels. *Toxicology*, 242(1-3):80-90.
- LaFranchi SH (2010). Newborn screening strategies for congenital hypothyroidism: an update. *Journal of Inherited Metabolic Disease*, 33(Suppl 2):S225-233.
- LaFranchi SH (2011). Increasing incidence of congenital hypothyroidism: some answers, more questions. *Journal of Clinical Endocrinology and Metabolism*, 96(8):2395-2397.
- LaFranchi SH, Austin J (2007). How should we be treating children with congenital hypothyroidism? *Journal of Pediatric Endocrinology and Metabolism*, 20(5):559-578.
- Lamm SH, Doemland M (1999). Has perchlorate in drinking water increased the rate of congenital hypothyroidism? *Journal of Occupational and Environmental Medicine*, 41(5):409-411.
- Larsen PR, Silva JE, Kaplan MM (1981). Relationships between circulating and intracellular thyroid hormones: physiological and clinical implications. *Endocrine Reviews*, 2(1):87-102.
- Lawrence J, Lamm S, Braverman LE (2001). Low dose perchlorate (3 mg daily) and thyroid function. *Thyroid*, 11(3):295.
- Lawrence JE, Lamm SH, Pino S, Richman K, Braverman LE (2000). The effect of short-term low-dose perchlorate on various aspects of thyroid function. *Thyroid*, 10(8):659-663.
- Leatherland JF, Sonstegard RA (1980a). Structure of thyroid and adrenal-glands in rats fed diets of Great-Lakes Coho Salmon (*Oncorhynchus-kisutch*). *Environmental Research*, 23(1):77-86.
- Leatherland JF, Sonstegard RA (1980b). Seasonal-changes in thyroid hyperplasia, serum thyroid-hormone and lipid concentrations, and pituitary-gland structure in Lake-Ontario Coho Salmon, *Oncorhynchus-kisutch walbaum* and a comparison with Coho Salmon from Lakes Michigan and Erie. *Journal of fish biology*, 16(5):539-562.
- Leatherland JF, Sonstegard RA (1982a). Bioaccumulation of Organochlorines by Yearling Coho Salmon (*Oncorhynchus-Kisutch Walbaum*) Fed Diets Containing Great-Lakes Coho Salmon, and the Pathophysiological Responses of the Recipients. *Comparative Biochemistry and Physiology. Part C, Pharmacology, Toxicology and Endocrinology*, 72(1):91-99.
- Leatherland JF, Sonstegard RA (1982b). Thyroid responses in rats fed diets formulated with Great-lakes Coho salmon. *Bulletin of Environmental Contamination and Toxicology*, 29(3):341-346.
- Lei J, Nowbar S, Mariash CN, Ingbar DH (2003). Thyroid hormone stimulates Na-K-ATPase activity and its plasma membrane insertion in rat alveolar epithelial cells. *American Journal of Physiology: Lung Cellular and Molecular Physiology*, 285(3):L762-772.
- Li FX, Byrd DM, Deyhle GM, Sesser DE, Skeels MR, Katkowsky SR, Lamm SH (2000a). Neonatal thyroid-stimulating hormone level and perchlorate in drinking water. *Teratology*, 62(6):429-431.
- Li Z, Li FX, Byrd D, Deyhle GM, Sesser DE, Skeels MR, Lamm SH (2000b). Neonatal thyroxine level and perchlorate in drinking water. *Journal of Occupational and Environmental Medicine*, 42(2):200-205.
- Longnecker MP, Gladen BC, Patterson DG, Jr., Rogan WJ (2000). Polychlorinated biphenyl (PCB) exposure in relation to thyroid hormone levels in neonates. *Epidemiology*, 11(3):249-254.
- Lyche JL, Oskam IC, Skaare JU, Reksen O, Sweeney T, Dahl E, Farstad W, Ropstad E (2004). Effects of gestational and lactational exposure to low doses of PCBs 126 and 153 on anterior pituitary and gonadal hormones and on puberty in female goats. *Reproductive Toxicology*, 19(1):87-95.
- Lyng GD, Snyder-Keller A, Seegal RF (2007). Polychlorinated biphenyl-induced neurotoxicity in organotypic cocultures of developing rat ventral mesencephalon and striatum. *Toxicological sciences : an official journal of the Society of Toxicology*, 97(1):128-139.

- McGrogan A, Seaman HE, Wright JW, de Vries CS (2008). The incidence of autoimmune thyroid disease: a systematic review of the literature. *Clinical Endocrinology*, 69(5):687-696.
- McNabb FM (2006). Avian thyroid development and adaptive plasticity. *General and Comparative Endocrinology*, 147(2):93-101.
- McNabb FMA (2005). Biomarkers for the assessment of avian thyroid disruption by chemical contaminants. *Avian and Poultry Biology Reviews*, 16(1):3-10.
- Meeker JD, Altshul L, Hauser R (2007). Serum PCBs, p,p'-DDE and HCB predict thyroid hormone levels in men. *Environmental Research*, 104(2):296-304.
- Mendelson CR, Boggaram V (1991). Hormonal control of the surfactant system in fetal lung. *Annual Review of Physiology*, 53:415-440.
- Mengreli C, Kanaka-Gantenbein C, Girginoudis P, Magiakou MA, Christakopoulou I, Giannoulia-Karantana A, Chrousos GP, Dacou-Voutetakis C (2010). Screening for congenital hypothyroidism: The significance of threshold limit in false-negative results. *Journal of Clinical Endocrinology and Metabolism*, 95(9):4283-4290.
- Miller MD, Crofton KM, Rice DC, Zoeller RT (2009). Thyroid-disrupting chemicals: interpreting upstream biomarkers of adverse outcomes. *Environmental Health Perspectives*, 117(7):1033-1041.
- Mitchell AM, Manley SW, Morris JC, Powell KA, Bergert ER, Mortimer RH (2001). Sodium iodide symporter (NIS) gene expression in human placenta. *Placenta*, 22(2-3):256-258.
- Mitchell ML, Hsu HW, Sahai I (2011). The increased incidence of congenital hypothyroidism: fact or fancy? *Clinical Endocrinology*, 75(6):806-810.
- Miyazaki W, Iwasaki T, Takeshita A, Tohyama C, Koibuchi N (2008). Identification of the functional domain of thyroid hormone receptor responsible for polychlorinated biphenyl-mediated suppression of its action in vitro. *Environmental Health Perspectives*, 116(9):1231-1236.
- Morreale de Escobar G, Obregon MJ, Escobar del Rey F (2000). Is neuropsychological development related to maternal hypothyroidism or to maternal hypothyroxinemia. *Journal of Clinical Endocrinology and Metabolism*, 85(11):3975-3987.
- Morris MS (2007). The association between serum thyroid-stimulating hormone in its reference range and bone status in postmenopausal American women. *Bone*, 40(4):1128-1134.
- Nicola JP, Basquin C, Portulano C, Reyna-Neyra A, Paroder M, Carrasco N (2009). The Na<sup>+</sup>/I<sup>-</sup> symporter mediates active iodide uptake in the intestine. *American Journal of Physiology. Cell Physiology*, 296(4):C654-662.
- National Research Council (2005). *Health implications of perchlorate ingestion*. Washington D.C.
- Oerbeck B, Sundet K, Kase BF, Heyerdahl S (2003). Congenital hypothyroidism: influence of disease severity and L-thyroxine treatment on intellectual, motor, and school-associated outcomes in young adults. *Pediatrics*, 112(4):923-930.
- Oerbeck B, Reinvang I, Sundet K, Heyerdahl S (2007). Young adults with severe congenital hypothyroidism: Cognitive event related potentials (ERPs) and the significance of an early start of thyroxine treatment. *Scandinavian Journal of Psychology*, 48(1):61-67.
- Olney RS, Grosse SD, Vogt RF (2010). Prevalence of congenital hypothyroidism-Current trends and future directions: Workshop summary. *Pediatrics*, 125:S31-S36.
- Oskam IC, Ropstad E, Smith AJ, Skaare JU, Tverdal A, Berg KA, Wiger R (2004). Effects of PCB99 and PCB153 exposure on spermatogenesis in young adult C57BL6 mice. *Reproductive Toxicology*, 19(2):169-180.
- Osman F, Gammage MD, Sheppard MC, Franklyn JA (2002). Clinical review 142: cardiac dysrhythmias and thyroid dysfunction: the hidden menace? *Journal of Clinical Endocrinology and Metabolism*, 87(3):963-967.
- Oxley JC, Smith JL, Higgins C, Bowden P, Moran J, Brady J, Aziz CE, Cox E (2009). Efficiency of perchlorate consumption in road flares, propellants and explosives. *Journal of Environmental Management*, 90(11):3629-3634.
- Panzica GC, Viglietti-Panzica C, Ottinger MA (2005). Introduction: neurobiological impact of environmental estrogens. *Brain Research Bulletin*, 65(3):187-191.
- Pearce EN, Spencer CA, Mestman JH, Lee RH, Bergoglio LM, Mereshian P, He X, Leung AM, Braverman LE (2011). The effect of environmental perchlorate on thyroid function in pregnant women from Cordoba, Argentina, and Los Angeles, California. *Endocrine Practice*:1-17.
- Pearce EN, Lazarus JH, Smyth PP, He X, Dall'amico D, Parkes AB, Burns R, Smith DF, Maina A, Bestwick JP, Jooman M, Leung AM, Braverman LE (2010a). Perchlorate and thiocyanate exposure and thyroid function in first-trimester pregnant women. *Journal of Clinical Endocrinology and Metabolism*, 95(7):3207-3715.
- Pearce MS, Korada M, Day J, Turner S, Allison D, Kibirige M, Cheetham TD (2010b). Increasing Incidence, but Lack of Seasonality, of Elevated TSH Levels, on Newborn Screening, in the North of England. *Journal of thyroid research*, 2010:101948.
- Persky V, Turyk M, Anderson HA, Hanrahan LP, Falk C, Steenport DN, Chatterton R, Jr., Freels S (2001). The effects of PCB exposure and fish consumption on endogenous hormones. *Environmental Health Perspectives*, 109(12):1275-1283.
- Pop VJ, Vulmsa T (2005). Maternal hypothyroxinaemia during (early) gestation. *Lancet*, 365(9471):1604-1606.
- Pop VJ, Brouwers EP, Vader HL, Vulmsa T, van Baar AL, de Vijlder JJ (2003). Maternal hypothyroxinaemia during early pregnancy and subsequent child development: a 3-year follow-up study. *Clinical Endocrinology*, 59(3):282-288.
- Pop VJ, de Vries E, van Baar AL, Waelkens JJ, de Rooy HA, Horsten M, Donkers MM, Komproe IH, van Son MM, Vader HL (1995). Maternal thyroid peroxidase antibodies during pregnancy: a marker of impaired child development? *Journal of Clinical Endocrinology and Metabolism*, 80(12):3561-3566.
- Razvi S, Ingoe L, Keeka G, Oates C, McMillan C, Weaver JU (2007). The beneficial effect of L-thyroxine on cardiovascular risk factors, endothelial function and quality of life in subclinical hypothyroidism: randomised, crossover trial. *Journal of Clinical Endocrinology and Metabolism*, 92(5):1715-1723.
- Rendon-Macias ME, Morales-Garcia I, Huerta-Hernandez E, Silva-Batalla A, Villasis-Keever MA (2008). Birth prevalence of congenital hypothyroidism in Mexico. *Paediatric and Perinatal Epidemiology*, 22(5): 478-485.
- Rodondi N, Aujesky D, Vittinghoff E, Cornuz J, Bauer DC (2006). Subclinical hypothyroidism and the risk of coronary heart disease: a meta-analysis. *The American journal of medicine*, 119(7):541-551.
- Roegge CS, Morris JR, Villareal S, Wang VC, Powers BE, Klintsova AY, Greenough WT, Pessah IN, Schantz SL (2006). Purkinje cell and cerebellar effects following developmental exposure to PCBs and/or MeHg. *Neurotoxicology and Teratology*, 28(1):74-85.
- Ropstad E, Oskam IC, Lyche JL, Larsen HJ, Lie E, Haave M, Dahl E, Wiger R, Skaare JU (2006). Endocrine disruption induced by organochlorines (OCs): Field studies and experimental models. *Journal of Toxicology and Environmental Health-Part a-Current Issues*, 69(1-2):53-76.
- Routti H, Nyman M, Jenssen BM, Backman C, Koistinen J, Gabrielsen GW (2008). Bone-related effects of contaminants in seals may be associated with vitamin D and thyroid hormones. *Environmental toxicology and chemistry / SETAC*, 27(4):873-880.
- Saita E, Hayama S, Kajigaya H, Yoneda K, Watanabe G, Taya K (2004). Histologic changes in thyroid glands from great cormorant (*Phalacrocorax carbo*) in Tokyo Bay, Japan: Possible association with environmental contaminants. *Journal of Wildlife Diseases*, 40(4):763-768.



- Sanchez CA, Barraji LM, Blount BC, Scrafford CG, Valentin-Blasini L, Smith KM, Krieger RI (2009). Perchlorate exposure from food crops produced in the lower Colorado River region. *Journal of Exposure Science and Environmental Epidemiology*, 19(4):359-368.
- Savu L, Vranckx R, Maya M, Nunez EA (1987). A thyroxine binding globulin (TBG)-like protein in the sera of developing and adult rats. *Biochemical and Biophysical Research Communications*, 148(3): 1165-1173.
- Savu L, Vranckx R, Maya M, Nunez EA (1989). Binding activities of thyroxine binding globulin versus thyroxine binding prealbumin in rat sera: differential modulation by thyroid hormone ligands, oleic acid and pharmacological drugs. *Biochemical and Biophysical Research Communications*, 159(3):919-926.
- Savu L, Vranckx R, Rouaze-Romet M, Maya M, Nunez EA, Treton J, Flink IL (1991). A senescence up-regulated protein: the rat thyroxine-binding globulin (TBG). *Biochimica et Biophysica Acta*, 1097(1):19-22.
- Scanes CG, McNabb FMA (2003). Avian models for research in toxicology and endocrine disruption. *Avian and Poultry Biology Reviews*, 14(1):21-52.
- Schantz SL, Widholm JJ, Rice DC (2003). Effects of PCB exposure on neuropsychological function in children. *Environmental Health Perspectives*, 111(3):357-576.
- Schell LM, Gallo MV, Denham M, Ravenscroft J, DeCaprio AP, Carpenter DO (2008). Relationship of thyroid hormone levels to levels of polychlorinated biphenyls, lead, p,p'- DDE, and other toxicants in Akwesasne Mohawk youth. *Environmental Health Perspectives*, 116(6):806-813.
- Schnitzler JG, Siebert U, Jepson PD, Beineke A, Jauniaux T, Bouquegneau JM, Das K (2008). Harbor porpoise thyroids: Histologic investigations and potential interactions with environmental factors. *Journal of Wildlife Diseases*, 44(4):888-901.
- Schuetz EG, Brimer C, Schuetz JD (1998). Environmental xenobiotics and the antihormones cyproterone acetate and spironolactone use the nuclear hormone pregnenolone X receptor to activate the CYP3A23 hormone response element. *Molecular Pharmacology*, 54(6):1113-1117.
- Seegal RF, Shain W (1992). Neurotoxicity of polychlorinated biphenyls: The role of ortho-substituted congeners in altering neurochemical function. In: (Isaacson RL, Jensen KF eds.) *The Vulnerable Brain and Environmental Risks*. New York, Plenum Press
- Shain W, Bush B, Seegal R (1991). Neurotoxicity of polychlorinated biphenyls: structure-activity relationship of individual congeners. *Toxicology and Applied Pharmacology*, 111(1):33-42.
- Sharlin DS, Tighe D, Gilbert ME, Zoeller RT (2008). The balance between oligodendrocyte and astrocyte production in major white matter tracts is linearly related to serum total thyroxine. *Endocrinology*, 149(5):2527-2536.
- Sharlin DS, Gilbert ME, Taylor MA, Ferguson DC, Zoeller RT (2010). The nature of the compensatory response to low thyroid hormone in the developing brain. *Journal of Neuroendocrinology*, 22(3):153-165.
- Skaare JU, Bernhoft A, Wiig O, Norum KR, Haug E, Eide DM, Derocher AE (2001). Relationships between plasma levels of organochlorines, retinol and thyroid hormones from polar bears (*Ursus maritimus*) at Svalbard. *Journal of Toxicology and Environmental Health. Part A*, 62(4):227-241.
- Smallridge RC, Glinoe D, Hollowell JG, Brent G (2005). Thyroid function inside and outside of pregnancy: what do we know and what don't we know? *Thyroid*, 15(1):54-59.
- Sonne C, Gustavson K, Riget FF, Dietz R, Birkved M, Letcher RJ, Bossi R, Vorkamp K, Born EW, Petersen G (2009). Reproductive performance in East Greenland polar bears (*Ursus maritimus*) may be affected by organohalogen contaminants as shown by physiologically-based pharmacokinetic (PBPK) modelling. *Chemosphere*, 77(11):1558-1568.
- Steinmaus C, Miller MD, Howd R (2007). Impact of smoking and thiocyanate on perchlorate and thyroid hormone associations in the 2001-2002 national health and nutrition examination survey. *Environmental Health Perspectives*, 115(9):1333-1338.
- Stoykov I, Zandieh-Doulabi B, Moorman AF, Christoffels V, Wiersinga WM, Bakker O (2006). Expression pattern and ontogenesis of thyroid hormone receptor isoforms in the mouse heart. *The Journal of endocrinology*, 189(2):231-245.
- Surks MI (1991). Thyroid-stimulating hormone: reference range validity. *JAMA: the journal of the American Medical Association*, 266(11):1573.
- Suvorov A, Takser L (2008). Facing the challenge of data transfer from animal models to humans: the case of persistent organohalogenes. *Environmental Health*, 7:58.
- Szinnai G, Lacroix L, Carre A, Guimiot F, Talbot M, Martinovic J, Delezoide AL, Vekemans M, Michiels S, Caillou B, Schlumberger M, Bidart JM, Polak M (2007). Sodium/Iodide Symporter (NIS) Gene Expression Is the Limiting Step for the Onset of Thyroid Function in the Human Fetus. *Journal of Clinical Endocrinology and Metabolism*, 92(1):70-76.
- Tabb MM, Kholodovych V, Grun F, Zhou C, Welsh WJ, Blumberg B (2004). Highly chlorinated PCBs inhibit the human xenobiotic response mediated by the steroid and xenobiotic receptor (SXR). *Environmental Health Perspectives*, 112(2):163-169.
- Takser L, Mergler D, Baldwin M, de Grosbois S, Smargiassi A, Lafond J (2005). Thyroid hormones in pregnancy in relation to environmental exposure to organochlorine compounds and mercury. *Environmental Health Perspectives*, 113(8):1039-1045.
- Tani Y, Mori Y, Miura Y, Okamoto H, Inagaki A, Saito H, Oiso Y (1994). Molecular cloning of the rat thyroxine-binding globulin gene and analysis of its promoter activity. *Endocrinology*, 135(6):2731-2736.
- Taurog A (2004). Hormone synthesis: Thyroid iodine metabolism. In: (Braverman LE, Utiger RD eds.) *The Thyroid: A Fundamental and Clinical Text*, Ninth edn., pp. 61-85. Philadelphia, Lippincott-Raven
- Tellez Tellez R, Michaud Chacon P, Reyes Abarca C, Blount BC, Van Landingham CB, Crump KS, Gibbs JP (2005). Long-term environmental exposure to perchlorate through drinking water and thyroid function during pregnancy and the neonatal period. *Thyroid*, 15(9):963-975.
- Turyk ME, Anderson HA, Persky VW (2007). Relationships of thyroid hormones with polychlorinated biphenyls, dioxins, furans, and DDE in adults. *Environmental Health Perspectives*, 115(8):1197-1203.
- Urbansky ET (2002). Perchlorate as an environmental contaminant. *Environ Sci Pollut Res Int*, 9(3):187-192.
- US EPA (2002). Perchlorate environmental contamination: Toxicological review and risk characterization. External review draft. (NCEA-1-0503 ed.). U.S. Environmental Protection Agency, National Center for Environmental Assessment, Office of Research and Development., Washington, D.C.
- Valentin-Blasini L, Blount BC, Otero-Santos S, Cao Y, Bernbaum JC, Rogan WJ (2011). Perchlorate exposure and dose estimates in infants. *Environmental Science and Technology*, 45(9):4127-4132.
- van den Hove MF, Beckers C, Devlieger H, de Zegher F, De Nayer P (1999). Hormone synthesis and storage in the thyroid of human preterm and term newborns: effect of thyroxine treatment. *Biochimie*, 81(5):563-570.
- van Tuyl M, Blommaert PE, de Boer PA, Wert SE, Ruijter JM, Islam S, Schnitzer J, Ellison AR, Tibboel D, Moorman AF, Lamers WH (2004). Prenatal exposure to thyroid hormone is necessary for normal postnatal development of murine heart and lungs. *Developmental Biology*, 272(1):104-117.
- Vanderpump MP (2011). The epidemiology of thyroid disease. *British Medical Bulletin*, 99:39-51.
- Vanderpump MPJ, Tunbridge WMG, French JM (1995). The incidence of thyroid disorders in the community: a twenty-year follow-up of the Wickham Survey. *Clinical Endocrinology*, 43(1):55-68.

- Vayre L, Sabourin JC, Caillou B, Ducreux M, Schlumberger M, Bidart JM (1999). Immunohistochemical analysis of Na<sup>+</sup>/I<sup>-</sup> symporter distribution in human extra-thyroidal tissues. *European Journal of Endocrinology*, 141(4):382-386.
- Vella KR, Hollenberg AN (2009). The ups and downs of thyrotropin-releasing hormone. *Endocrinology*, 150(5):2021-2023.
- Verreault J, Skaare JU, Jenssen BM, Gabrielsen GW (2004). Effects of organochlorine contaminants on thyroid hormone levels in arctic breeding glaucous gulls, *Larus hyperboreus*. *Environmental Health Perspectives*, 112(5):532-537.
- Vranckx R, Savu L, Nunez EA (1989). The microheterogeneity of rat TBG. *FEBS Letters*, 244(2):343-346.
- Vranckx R, Rouaze-Romet M, Savu L, Mechighel P, Maya M, Nunez EA (1994). Regulation of rat thyroxine-binding globulin and transthyretin: studies in thyroidectomized and hypophysectomized rats given tri-iodothyronine or/and growth hormone. *The Journal of endocrinology*, 142(1):77-84.
- Wan W, Farhoud B, Privalsky ML (2005). Pituitary resistance to thyroid hormone syndrome is associated with T3 receptor mutants that selectively impair beta2 isoform function. *Molecular Endocrinology*, 19(6):1529-1542.
- Whitfield GK, Jurutka PW, Haussler CA, Haussler MR (1999). Steroid hormone receptors: evolution, ligands, and molecular basis of biologic function. *Journal of Cellular Biochemistry*, Suppl 32-33:110-122.
- Wolff J (1998). Perchlorate and the thyroid gland. *Pharmacological Reviews*, 50(1):89-105.
- Wu W, Niles EG, LoVerde PT (2007). Thyroid hormone receptor orthologues from invertebrate species with emphasis on *Schistosoma mansoni*. *BMC Evolutionary Biology*, 7:150.
- Yamano K, Araki K, Sekikawa K, Inui Y (1994). Cloning of thyroid hormone receptor genes expressed in metamorphosing flounder. *Developmental Genetics*, 15(4):378-382.
- You SH, Gauger KJ, Bansal R, Zoeller RT (2006). 4-Hydroxy-PCB106 acts as a direct thyroid hormone receptor agonist in rat GH3 cells. *Molecular and Cellular Endocrinology*, 257-258:26-34.
- Zarkovic M, Ciric J, Beleslin B, Ciric S, Bulat P, Topalov D, Trbojevic B (2011). Further studies on delineating thyroid-stimulating hormone (TSH) reference range. *Hormone and Metabolic Research*, 43(13):970-976.
- Zhou T, John-Alder HB, Weis JS, Weis P (2000). Endocrine disruption: thyroid dysfunction in mummichogs (*Fundulus heteroclitus*) from a polluted habitat. *Marine Environmental Research*, 50(1-5):393-397.
- Zhou T, Ross DG, DeVito MJ, Crofton KM (2001). Effects of short-term in vivo exposure to polybrominated diphenyl ethers on thyroid hormones and hepatic enzyme activities in weanling rats. *Toxicological sciences : an official journal of the Society of Toxicology*, 61(1):76-82.
- Zhou T, Taylor MM, DeVito MJ, Crofton KM (2002). Developmental exposure to brominated diphenyl ethers results in thyroid hormone disruption. *Toxicological sciences : an official journal of the Society of Toxicology*, 66(1):105-116.
- Zimmermann MB (2007). The adverse effects of mild-to-moderate iodine deficiency during pregnancy and childhood: a review. *Thyroid*, 17(9):829-835.
- Zoeller RT (2001). Polychlorinated biphenyls as disruptors of thyroid hormone action. In:(Fisher LJ, Hansen L eds.) *PCBs: Recent Advances in the Environmental Toxicology and Health Effects of PCBs*, pp. 265-272. Lexington, University of Kentucky Press
- Zoeller RT (2003). Thyroid toxicology and brain development: should we think differently? *Environmental Health Perspectives*, 111(12):A628.
- Zoeller RT (2010). Environmental chemicals targeting thyroid. *Hormones (Athens)*, 9(1):28-40.
- Zoeller RT, Rovet J (2004). Timing of thyroid hormone action in the developing brain: clinical observations and experimental findings. *Journal of Neuroendocrinology*, 16(10):809-818.
- Zoeller RT, Tan SW, Tyl RW (2007a). General background on the hypothalamic-pituitary-thyroid (HPT) axis. *Critical Reviews in Toxicology*, 37(1):11-53.
- Zoeller RT, Tyl RW, Tan SW (2007b). Current and potential rodent screens and tests for thyroid toxicants. *Critical Reviews in Toxicology*, 37(1):55-95.